Check for updates

The protein corona from nanomedicine to environmental science

Morteza Mahmoudi 🖗 1 🖂, Markita P. Landry 🕲 ^{2,3,4,5}, Anna Moore¹ & Roxana Coreas 🕲 ² 🖂

Abstract

The protein corona spontaneously develops and evolves on the surface of nanoscale materials when they are exposed to biological environments, altering their physiochemical properties and affecting their subsequent interactions with biosystems. In this Review, we provide an overview of the current state of protein corona research in nanomedicine. We next discuss remaining challenges in the research methodology and characterization of the protein corona that slow the development of nanoparticle therapeutics and diagnostics, and we address how artificial intelligence can advance protein corona research as a complement to experimental research efforts. We then review emerging opportunities provided by the protein corona to address major issues in healthcare and environmental sciences. This Review details how mechanistic insights into nanoparticle protein corona formation can broadly address unmet clinical and environmental needs, as well as enhance the safety and efficacy of nanobiotechnology products.

Sections

Introduction

Identity of the protein corona for nanomedicine

Challenges in the research methodology and characterization of the protein corona

Emerging opportunities offered by the protein corona

Conclusion and perspectives

¹Department of Radiology and Precision Health Program, Michigan State University, East Lansing, MI, USA. ²Department of Chemical and Biomolecular Engineering, University of California, Berkeley, CA, USA. ³Innovative Genomics Institute, Berkeley, CA, USA. ⁴California Institute for Quantitative Biosciences, University of California, Berkeley, CA, USA. ⁵Chan Zuckerberg Biohub, San Francisco, CA, USA. ^{Ser}email: mahmou22@msu.edu; roxanacoreas@berkeley.edu

Introduction

Nanoparticles (NPs) are increasingly prevalent in our society, with applications ranging from nanomedicine, photovoltaics, cosmetics and beyond. Despite the ubiquity of their use, we still lack a fundamental understanding of how NPs interact with their local environments. This poor understanding becomes a critical problem when using NPs in real-life situations. Notably, it has been well documented that introducing NPs into biological fluids results in the formation of a surrogated biological identity known as the biomolecular corona, which is the spontaneous adsorption of proteins¹, lipids², sugar moieties³, nucleic acids⁴ and metabolites^{5,6} onto nanomaterial surfaces. The composition of the biomolecular corona strongly determines the ultimate fate of NPs⁷⁻⁹. As most studies that address the biomolecular corona formation have focused on proteins adsorbed to the surface of NPs¹⁰, together with the fact that the predominant model for the formation of the biomolecular corona is hegemonized by the direct adsorption of proteins from biological milieus¹¹, this Review comprehensively focuses on the protein corona.

The emergence of the concept of the protein corona in nanomedicine substantially altered perspectives of researchers and interpretation of biosystem-NP interactions. The first critical milestone occurred in 2007 when Dawson's group conceived the term protein corona¹². However, protein corona studies commenced as early as the late 1950s¹³ and early 1960s^{14,15} under a slightly different terminology and a colloidal science context. The characterization of protein coronas has revealed that the adsorption of proteins onto nanomaterials can have mixed biological effects. Protein corona patterns, which are dependent on the physicochemical properties of nanomaterials and the complexities of biological matrices, can unpredictably change NP outcomes including function¹⁶, uptake^{17,18}, biodistribution^{19,20}, immunological responses^{10,21} and toxicity^{22,23}, and can thus be a challenge for therapeutic nanomedicine. Mechanistically, the protein corona can be composed of endogenous ligands that can mask the innate or designed reactivity of NP surfaces, block cell membrane receptors thus hindering NP internalization, protect NPs from opsonization (that is, the immunological response that targets NPs for removal through the adsorption of antibodies and complement proteins onto the surfaces of NPs), limit blood circulation or enhance cytotoxicity compared with their pristine NP counterparts.

Juxtaposingly, the protein corona also creates new opportunities for diagnostic and personalized nanomedicine; the protein corona can be utilized for disease diagnosis^{24–27}, for the optimization of cell internalization²⁸⁻³⁰ or leveraged for improving the in vivo biodistribution^{21,31} of nanomedicines. The enrichment of plasma proteins, particularly rare proteins and glycoproteins, onto NPs can create a protein corona 'fingerprint', which can be used for the personalized detection of biomarkers and assist in post-translational modifications in support of risk stratification, prognosis and disease recognition. Selectively coating NPs with purposely designed protein coronas can regulate cell-dependent uptake, promote blood circulation and enhance their therapeutic efficacies. Furthermore, protein corona studies have advanced from the conceptualization on how biosystems 'see' and perceive NPs³² towards the development of new diagnostics and therapies^{24-26,33,34}, the emergence of the eco-corona³⁵⁻³⁷ and materialization of the biological and ecological impacts induced by protein-corona-coated nanoplastics^{38,39}. Artificial intelligence (AI) can be harnessed for data-driven discoveries of NP-biological interactions and complexities. Machine learning algorithms can identify the important variables that affect protein corona formation on specific types of NPs⁴⁰ and have predicted diseases in patients using personalized protein corona fingerprints^{19,26}. Accordingly, a better understanding of the composition, pattern and decoration of biomolecules at the surface of NPs, supplemented by AI, can facilitate the development of safer and more effective nanomedicine technologies with desired biological fates. Several challenges remain in the field of protein corona research, including NP heterogeneity, interpretation of protein patterns and induced perturbations of immunological and toxicological responses. Through standardization of methodologies and detailed characterization of the protein corona, foundational for data sets that fuel AI, targeted therapeutic and/or diagnostic nanomedicines can be optimized.

In this Review, we summarize the progress, challenges and opportunities in protein corona research. We expose the current state of protein corona research in nanomedicine, highlight current challenges in research methodology and characterization and address how AI can be used to tackle these challenges. We then discuss emerging opportunities offered by the protein corona for the design of efficient nanomedicines and innovative therapies, in proteomicbased screening and for the detection of diseases. We also address the importance of understanding the protein corona to assess the ecotoxicological impacts of nanomaterials that traverse through the environment.

Identity of the protein corona for nanomedicine Understanding the biofluid protein corona

The scientific understanding of the protein corona for nanomedicine applications began with empirical analyses of the as-of-yet uncharacterized protein corona that formed when polystyrene NPs interacted with proteins in complex biological fluids, such as plasma or serum⁴¹⁻⁴³. These studies evaluated the efficiency of polymers to hinder protein adsorption onto colloidal particles, with the goal of enhancing drug targeting and correlating adsorbed protein patterns with the role of surface charges on blood cells. Thus, although the protein corona was inadvertently analysed, the main aim was not its characterization and exploitation. Following these studies, protein conformation⁴⁶⁻⁴⁸ and the functionality of the adsorbed proteins on the surfaces of NPs⁴⁹⁻⁵¹. Figure 1 provides a timeline of key milestones within protein corona research.

Equilibrium binding between proteins and NPs is largely dependent on protein identity and on NP physicochemical properties (such as size and surface chemistry)^{12,44}. Binding affinities for the adsorption and desorption of proteins affect the colloidal stability of NPs. Proteins may also change conformation upon adsorption onto NPs as conformational changes are dependent on both protein and NP properties. Greater conformational changes are observed in proteins adsorbed onto hydrophobic NP surfaces relative to their hydrophilic counterparts, with the shape of the protein impacting its stability on NPs with high surface curvatures^{46,47}. External factors such as pH and ionic strength also influence surface-binding-induced protein unfolding⁴⁵. For example, adsorption-induced protein conformational changes were quantified by targeting and fluorescently labelling exposed amine groups; these residues, which were originally buried inside the tertiary structures of proteins in native conformations, became exposed because of adsorption-induced conformational disorders⁴⁸. Protein conformational changes can impact protein functionality through the exposure of once-concealed functional epitopes or misfolding that results in complete loss-of-protein function, subsequently impacting



downstream biological processes and mediating NP-biosystem interactions.

By studying the spatial organization of the protein corona on polystyrene NPs, the location of adsorbed proteins was discovered to be randomly distributed. Yet, these randomly adsorbed proteins possessed functional epitopes that could bind to receptors⁴⁹. Similarly, adsorbed lipoproteins on 2D graphene flakes were found to possess several functional epitopes capable of binding to receptors in the liver⁵⁰. Furthermore, analysis of adsorbed low-density lipoproteins and IgG on SiO₂ NPs revealed functional epitopes, which enhanced NP uptake in human lung and embryonic kidney cells⁵¹. Overall, the functionality of the protein corona, which is influenced by nanomaterial physiochemical properties, protein varieties and concentrations in biofluids and ambient parameters (such as pH and temperature), allows for the specific recognition of cell receptors and suggests that there may exist multivalent interactions between cells and NPs mediated by rare corona proteins that are, to date, not well understood.

The understanding of corona-mediated functionalities can also be leveraged for the development of nanomedicines. Personalized protein^{27,52-54} and biomolecular coronas^{4-6,55} have been proposed as tools for disease diagnosis and prognosis, as the composition of the protein corona developed in patient-derived serum is disease dependent. For instance, the protein corona formed in patient sera can be used to identify biomarkers for lung cancer⁵², predict Alzheimer disease⁵³ and screen for pancreatic cancer⁵⁴. Multi-omics of the biomolecular corona^{4,5,55}, which combines the analyses of metabolites, including organic acids, sugars, amino acids and hormones, as well as lipids and proteins, has enabled a more thorough profiling of human samples for disease diagnostics, but these studies are limited by extraction methods and separation conditions. Methodical sample preparation is crucial and must account for surface chemistry, along with pH and ionic strength of elution and/or extraction buffers, to isolate metabolite, protein and lipid constituents from the biomolecular corona⁶. Additionally, protein coronas have been utilized to maximize the performance of traditional proteomic pipelines by deep sampling complex plasma proteomes, comprising thousands of proteins in human plasma samples²⁶. Traditional proteomic analysis of the plasma proteome has been difficult to achieve owing to the presence of a few dozens of extremely abundant proteins that dominate the protein mass content in plasma. Formation of the protein corona on NPs results in the enrichment of rare proteins and biomarkers within the proteome that can be identified by liquid chromatography-mass spectrometry

(LC–MS/MS)^{24–27}. Recently, protein coronas that formed on five different NPs were utilized to systematically detect over 2,000 proteins from more than 100 plasma samples, bypassing the typical complex sample preparation workflows for neat plasma that immunodeplete highly abundant proteins and fractionate samples with chromatography. These protein corona data sets were then utilized to predict low-abundance proteins associated with non-small-cell lung cancer²⁶.

Soft and hard coronas

NPs possess unique physiochemical properties⁵⁶, such as size^{57,58}, shape⁵⁹ and surface chemistries^{19,57,60}, that can affect the formation of the protein corona. Additionally, the corona profile can be influenced by ambient factors such as biomolecule concentration⁶⁰, pH⁶¹, ionic strength⁶², temperature⁶³ and incubation time^{64–66}. Upon corona formation, the identity of the NP is transformed; its physicochemical properties⁶⁷, targeting abilities^{20,28,68} and biological responses^{17,22,65,69} change owing to the surrogated biological identity dictated by the adsorbed proteins. For example, characterization of the protein corona on core-shell NPs, composed of identical PEGylated lipid bilayer shells with varying core elasticities, revealed that apolipoprotein A1 preferentially adsorbs onto NPs with intermediate elasticity and the adsorption strongly correlates with longer NP elasticity-dependent in vivo systemic circulation lifetimes⁷⁰.

Time-dependent profiling of protein coronas, extracted from human plasma, revealed that the formation of protein corona fingerprints occurs rapidly (-30 s), and the amount of protein in the corona, but not the composition of the protein profile, can change over time⁷¹. Moreover, the rapid formation of the protein corona impacts the kinetic pathophysiology of the NPs. For example, both an increased NP uptake by microvascular endothelial cells and the inhibition of erythrocyte haemolysis were observed upon protein corona formation⁷¹. The study of in situ protein corona formation in complex biological matrices, such as whole blood and plasma, showed that PEGylated gold NPs do not aggregate in these fluidic environments⁷².

Protein coronas incorporate both soft and hard coronas. The soft corona, comprising biomolecules such as proteins with low affinities to NP surfaces, endures dynamic and reversible exchanges that are dependent on the conditions of the surrounding biological fluid^{58,73,74}. The hard corona develops gradually, is more stable and is composed of 'relatively immobile' proteins with substantial affinities to, and low tendencies to dissociate from, NP surfaces^{57,60,64}. The understanding of both soft and hard coronas is crucial for inferring NP stability, function

and interactions with biological systems. Kinetically, the associations between proteins and NPs in biological fluids are governed by noncovalent interactions, such as electrostatic forces, hydrophobic forces, hydrogen bonding and $\pi - \pi$ stacking⁷⁵. Proteins competitively bind to NP surfaces⁷⁶ establishing transient NP-protein complexes composed of soft and hard corona proteins^{58,64,73,74} under thermodynamically favourable conditions⁷⁵. Because of the high dissociation rate of the soft corona proteins, our current understanding of the biological identity of the protein corona is typically limited to the hard corona proteins. Moreover, both NP properties and the conditions of biological matrices can impact the kinetic and dynamic binding equilibrium of the soft and hard protein corona to NP surfaces. Protein-protein interactions on the hard protein corona can either be stable or transient similar to the soft corona⁷⁷. The majority of soft corona proteins are not solely unique to the soft corona composition, but rather also compose the hard corona profile, indicating that soft corona proteins possess variable binding strength states⁷⁸. Moreover, artificially hardening the dynamic soft corona to increase the residence time of the soft corona proteins revealed decreased cellular associations, suggesting that the dissociation of soft corona proteins can reveal bare NP surfaces allowing for nonspecific interactions with cell membranes78.

Activation of bioorthogonal nanozymes, in this case transition metal catalysts that mimic enzymes, varies depending on the dynamic nature of the protein corona: ex vivo, the soft corona reduces nanozyme functionality, but the hard corona results in aggregation and total loss of nanozyme activity; intracellularly, endosomal and lysosomal proteases restore the catalytic activity of both the hard and soft corona-coated nanozymes¹⁶. Protein corona formation can also be stereo-specific. For example, the composition of the soft corona of chiral cuprous sulfide (Cu₂S) is chirality-dependent and correlates to its stereo-selective biodistribution⁷⁹. Time-dependent changes to the chirality-mediated soft and hard corona compositions were correlated with decreased blood circulation and trafficking to the liver⁷⁹. Silver NPs typically dissolve into biologically active and highly toxic Ag^+ ions⁸⁰. However, the presence of a hard corona on the NPs induces their sulfidation, which decreases their toxicity by forming insoluble Ag₂S nanocrystals⁸¹; meanwhile, the soft corona was found to mediate Ag^+ removal, reducing Ag_2S nanocrystal formation⁸². These previous studies highlight that to better comprehend the distinct biological responses of nanomedicines, the differential compositions of the soft and hard protein coronas need to be taken into consideration.

Remaining challenges limiting nanomedicine development

There remain several challenges within the protein corona discipline that must be taken into consideration to improve nanomedicines.

Protein patterns. Although several studies have identified the composition and abundances of adsorbed proteins on NP surfaces, few have been able to distinguish protein patterns, such as multifaceted protein recognition domains, and their roles towards NP recognition pathways by biosystems³². Although NPs coated with protein coronas can interact with cell membranes directly through ligand–receptor connections, transient interactions with multiple proteins on the corona can also occur on the membrane. It is possible that the presence of multiple membrane receptors can facilitate the interaction with multiple functional epitopes exposed on the protein corona. These short-lived interfaces can recruit intracellular biomolecules that form cytosolic signal transduction clusters and ultimately bionanosynapse with protein corona-coated NPs, thus preparing cells for ensuing biological

impacts³². Moreover, there is a knowledge gap in distinguishing all of the multivalent roles of corona proteins with regard to their recognition by cells, in NP pharmacokinetics as well as immunoregulatory signalling and gene expression. Decoding which corona protein pattern impacts cell internalization, biological distribution into tissues and organs and the secretion of inflammatory cytokines and genetic material is an extraordinarily challenging question.

Differing biological and toxicological responses. The identity and nature of the adsorbed corona proteins can mediate biological and toxicological responses; even though the protein corona can provide a protective coating and mitigate cytotoxicity⁸³⁻⁸⁵, it also prompts enhanced toxicity in some cases⁸⁶⁻⁸⁸. Although there exist several differences in protein corona profiles (such as the presence of rare proteins in some cases) extracted from NP surfaces, multiple studies have confirmed that abundant proteins - such as albumin, fibrinogen, apolipoproteins, complement proteins, transferrin and immunoglobulins - commonly adsorb on NPs because of their relatively high abundance in human serum and plasma⁸⁹. These adsorbed proteins can influence NP physiological responses. For example, although bare carboxylated-multiwalled carbon nanotubes (CNTs) induced blood platelet aggregation and release of platelet membrane microparticles, coating the CNTs with albumin and fibrinogen attenuated platelet aggregation and prevented the release of membrane microparticles, respectively⁹⁰.

Protein conformational changes induced by adsorption can also mediate biological responses. For example, albumin changed conformation following its adsorption onto nanoporous polymer NPs (NPP), with cell-dependent differential uptake mediated by the unfolded albumin. In that case, a substantial decrease in NPP uptake by monocytes, but a slightly elevated receptor-mediated phagocytosis of NPP in macrophages, was observed⁹¹. Similarly, conformational changes of fibrinogen upon adsorption onto gold NPs induced the activation of cell receptors, upregulated transcription factor signalling and released inflammatory cytokines in monocytes⁶⁹. Conformational changes to IgG, but not to fibrinogen, upon adsorption to CNTs were associated with elevated levels of reactive oxygen species and inflammatory cytokines from macrophages, mediated by the denatured IgG adsorbed on CNTs⁹². Currently, reports describing the nature of the adsorbed protein corona and subsequent paradoxical biological impacts are limited because of the lack in diversity of protein-NP systems under investigation and because of deficiencies in understanding protein-NP adsorption mechanisms93, thus precluding generalizable trends of protein corona dynamics.

Perturbed immunological response. The injection of nanomedicines into the circulatory system can result in perturbed immunological responses as well as the unsolicited formation of the protein corona, opsonization or immune system recognition of specific corona patterns that limit the circulation of nanotherapeutics^{65,94,95}. For example, during the opsonization of superparamagnetic iron oxide nanoworms in human plasma, complement component 3 (C3) covalently bound to absorbed proteins at the surface of the used magnetic nanoworms. C3, which is the most abundant complement protein in serum, activates an immunological response (the complement system) for the removal of foreign materials, such as NPs, from cells. After the binding of C3 to the nanoworm protein corona in vitro, a dynamic exchange was observed in vivo, suggesting that the immunological corona was kinetically unstable²¹. The exchangeable nature of the protein corona may

induce re-recognition by the immune system in vivo as it does in vitro²¹, resulting in rapid, undesirable clearance of nanomedicines^{94,95}. Further physiologically relevant in vivo protein corona exploration is necessary to engineer nanomedicines that deter immunological responses, such as complement protein exchange, and enhance circulation time.

Nanoparticle heterogeneity. Separation of heterogeneous NPs and analysis of the individual corona from a single particle is a challenging task that has not been studied extensively^{96,97}. The MagLev (magnetic levitation) technique, which uses a high-intensity magnetic field to levitate and separate diamagnetic objects according to their densities, has been applied to study protein corona heterogeneity and has been suggested for extraction of homogeneous corona-coated NPs%. Future endeavours should conduct global analyses of the variances in protein corona, as typical 'averages' of the protein corona do not constitute all corona subclasses⁹⁷, just as single-cell gene expression analysis can have striking differences from the averaged analysis of a population of cells⁹⁸. Thus, there are several reasons why it is crucial to characterize NP protein coronas and analyse their biological impacts for the development of reproducible nanomedicines⁹⁹. Integration of high-throughput computational tools, such as AI, can complement high-throughput protein corona investigations to catalyse the advancement of nanomedicine.

Targeting strategies for therapeutic nanomedicine

NPs that have targeting capabilities are obtained by functionalizing their surface with targeting species (such as antibodies, small molecules and nucleic acids), enabling their localization to desired locations in the body for payload release and/or imaging purposes. However, the formation of the protein corona can shield these targeting moieties and cause mistargeting and unfavourable biodistribution²⁰.

There are four major proposed strategies to address the shielding role of the protein corona on targeted NPs (Fig. 2).

Protein-repellent coating. The first strategy involves the use of protein-repellent coatings (such as zwitterionic compounds) on the surface of NPs to prevent and/or minimize corona formation^{100,101} (Fig. 2a). For example, silica NPs functionalized with cysteine (as a zwitterionic ligand) and conjugated with biotin (as a targeting molecule) prevented the formation of the protein corona, when compared with the same NP without cysteine functionalization¹⁰¹. The zwitterionic coating also substantially improved the targeting capacity. Another relevant example is the use of antifouling polymers (such as polyethylene glycol or polyethylene oxide)¹⁰², which reduce shielding by the protein corona on the basis of the characteristics of the specific coating (such as density, size, length and heterogeneity of the polymeric coating)^{103,104}.

Pre-coating strategy. Another strategy is to pre-coat NPs with components that recruit plasma proteins with targeting capacities during corona formation²⁹ (Fig. 2b). The pre-coating strategy, which was first used in 1980 to inhibit uptake of liposomes by macrophages¹⁰⁵, can be precisely designed to manipulate protein corona composition in a way that provides a new impetus for targeted payload delivery to specific biosystems and/or tissues. For example, surface modification of liposomes with a short amyloid β -derived peptide can recruit plasma apolipoproteins, and the resulting apolipoprotein corona-coated liposomes can target brain tissue for high-yield drug delivery applications¹⁰⁶. One of the main challenges of the pre-coating approach, however, is that the active site of proteins should be exposed in the outer

corona layer for interaction with specific cell receptors. For instance, although silica NPs pre-coated with the serum proteins y-globulins could enrich the protein corona with immunoglobulins and opsonin, which have high targeting efficacy and recognition to macrophages through Fc (fragment crystallizable) receptors, there was no substantial enhancement of uptake of these NPs by macrophages compared with non-pre-coated NPs²⁹. This result highlights the importance of exposing the functional binding motifs to cell receptors⁴⁹, which is difficult to engineer from the spontaneous adsorption of such proteins to the NP surface. Therefore, new strategies should be developed to increase the efficiency of the pre-coating approach by enhancing the availability of functional motifs of corona proteins for cell receptors to achieve desired targeting efficacy⁴⁹. We now understand that precoating with albumin can bring more carrier proteins to the surface of NPs and improve their blood circulation time, therefore improving their biodistribution profile and targeting efficacies.

Pre-adsorption versus chemical conjugation. The third strategy entails using a physical approach to attach targeting moieties to the surface of NPs rather than chemical conjugation¹⁰⁷ (Fig. 2c). For instance, two types of targeted polystyrene NPs were synthesized by pre-adsorption versus chemical conjugation of anti-CD63 antibodies to their surfaces, and their targeting efficacies towards monocyte-derived dendritic cells with CD63 surface receptors, in the presence and absence of serum or plasma, were studied¹⁰⁷. Although, in the absence of serum or plasma, both targeted NPs demonstrated similar targeting capacity towards dendritic cells, the pre-absorbed antibody-coated NPs showed substantially higher targeting efficacy compared with chemically attached targeted NPs in the presence of serum or plasma. This is, at least in large part, because the pre-adsorption method allows attachment of antibodies with outward-facing recognition domains and better targeting outcomes compared with chemical conjugation.

Conjugation to protein corona. The last strategy involves attaching targeting moieties to the surface of corona-coated NPs¹⁰⁸ (Fig. 2d). The targeting species can be attached to the formed protein corona in an equilibrium state – when the protein corona is formed within a controlled single-component or multicomponent biofluid that does not change over time. For example, both bare and corona-coated gold and silica NPs were functionalized with transferrin, and their targeting capacities towards cells with transferrin receptors were monitored in the absence and presence of serum proteins¹⁰⁸. Although the targeting capacity of transferrin-conjugated bare gold and silica NPs in the presence of serum was reduced compared with the serum-free media, there was no evidence of targeting efficacy reduction when transferrin was conjugated to the equilibrated protein corona of gold and silica NPs.

Challenges in the research methodology and characterization of the protein corona Methodology

Poor methodology can induce errors and/or lead to misinterpretation of NP protein corona outcomes¹⁰⁹. Here, we focus on how the formation of the protein corona may cause misinterpretation in well-defined biological methodologies including toxicity assays and monitoring drug release strategies.

The interference of protein corona with cytotoxicity assays (including the 3-[4,5-dimethylthiazol-2yl]-2,5-diphenyltetrazolium bromide (MTT) assay) was one of the first issues raised¹¹⁰. The formation of the protein corona can substantially change the composition



Fig. 2 | Schematics showing the major challenges created by the NP protein corona and proposed strategies to address them. a, The use of proteinrepellent compounds to coat nanoparticles (NPs) can minimize corona formation and, therefore, reduce the shielding effect of corona on active/ functional sites of targeting moieties. b, The use of specific proteins to pre-coat NPs to enhance recruitment of proteins with intrinsic targeting capacities during corona formation. c, Maintaining the exposure of active site of antibodies, even after corona formation by pre-adsorbing targeting antibodies to the surface of NPs. d, Using strategies to attach targeting moieties to the surface of corona-coated NPs.

and nutritional balance of cell culture media in static settings mainly because of the attraction of proteins, amino acids and vitamins to the surface of NPs and because of variations in protein conformation after interactions with the surface of NPs. As a result, the modified cell culture medium itself can induce cytotoxic effects under static in vitro conditions and may cause errors in NP toxicity outcomes. Two main strategies can be used to achieve reliable NP toxicological data. The first approach is to use a bioreactor, in which the use of dynamic flow can minimize the protein-NP adsorptive effects in cell culture media, thereby avoiding NP-induced variabilities in media composition. The second strategy is to introduce NPs to the cell culture media first and incubate them for an hour (to ensure that equilibrium among the biomolecules on the NP surface is reached) and then collect the particles and introduce them to cells in fresh culture media to minimize the effect of the protein corona on the composition of the culture media.

Interactions of NPs with proteins whose oligomerizations and fibrillations can cause neurodegenerative diseases (such as amyloid β -synuclein and α -synuclein for Alzheimer and Parkinson diseases, respectively) have been a subject of extensive research in nanomedicine¹¹¹⁻¹¹³. However, the role of NPs in the fibrillation process has often been probed in the absence of the protein corona. As NPs that reach brain tissues for possible interactions with amyloid or synuclein proteins would certainly have previously interacted with biological fluids, their interaction with neurodegenerative-related proteins should

also be studied in the presence of the protein corona. Corona-coated NPs mediate the antifibrillation impact of NPs and slow amyloid β -fibrillation¹¹⁴, which may cause a challenge in clinical translation of neurodegenerative nanotechnologies: ignoring the critical role of protein corona in probing the role of NPs in the fibrillation process can cause substantial misinterpretation of the outcomes, which, in turn, can lead to incorrect prediction of behaviour of NPs in vivo. Therefore, to achieve robust and clinically relevant effects on neurodegeneration-related proteins, corona-coated NPs instead of bare NPs should be studied.

Another major field in which the formation of the protein corona can make a substantial impact is the development of drug delivery nanoplatforms¹¹⁵. The protein corona can create an additional barrier to payload release, substantially influencing payload release profiles and even mechanism of action^{116,117}. As a result, the reported outcomes of payload release studies that lack consideration of the protein corona may not accurately represent the in vivo payload release from the nanoplatform. Corona formation can attenuate or accelerate drug release kinetics on the basis of the physicochemical properties of a nanocarrier, the corona itself and/or influential parameters (such as pH and temperature). Although studies have begun to consider the critical role of protein corona in the payload release kinetics of nanocarriers¹¹⁷, the effects on release kinetics of stimulus-responsive nanocarriers are poorly understood. Future studies should focus on the role of the protein corona on nanocarriers that respond to exogenous and/or

physiological stimulus including light, electric field, magnetic field, temperature and pH.

Overall, a deeper understanding of the methodological implications of the protein corona in various critical assays (such as toxicity and payload release) in nanomedicine would enable scientists and drug developers to design safe and more predictably efficient therapeutic nanomedicine technologies.

Characterization

The robust and precise characterization of the physicochemical properties and colloidal stability of corona-coated NPs are crucial for the identification of possible protein contamination and for the interpretation of protein corona outcomes¹¹⁸. Here, we focus on characterizing the composition of the protein corona in terms of protein identity and abundance, which is key to predicting and interpreting the interactions of NPs with biosystems. LC–MS/MS is one of the few techniques being used to define the type and abundance of proteins in the NP corona layer. Therefore, understanding the complexity of LC–MS/MS, from sample preparation methodologies to data analysis, is essential to accurately predict the biological fate of NPs.

Many laboratories that study protein corona have expertise in characterizing and analysing the physicochemical properties of the protein corona in house, but they are not necessarily specialized in MS-based proteomics; therefore, they rely on core facilities for proteomic analysis of their samples. As different MS-based proteomics laboratories and/or core facilities may use different methods in their LC-MS/MS workflow, different instruments, equipment and commercial software, one can expect that the heterogenicity of the outcomes would be substantial^{99,119-122}. To shed more light on how much and to what extent LC-MS/MS characterization and analysis can affect protein corona outcomes, 17 identical aliquots of corona-coated polystyrene NPs were sent to 17 different laboratories/core facilities for proteomics analysis¹²³. The outcomes were surprising: out of 4,022 identified unique proteins in the protein corona layer, only 73 (1.8%) were shared across the laboratories and/or core facilities. It is noteworthy that the technical repeats from each of the core facilities revealed reproducible results, which emphasizes that using the identical sample preparation approach and instrumentation can provide reliable results. The observed heterogeneity across laboratories and/or core facilities, however, is an extremely important point, which needs to be seriously considered in nanomedicine literature, as any interpretation regarding the interactions of NPs with biological systems heavily relies on the composition of protein corona. To improve the reliability and robustness of protein corona data, the nanobio interface community should develop standard protocols on methodologies, analysis, reporting and interpretation of LC-MS/MS data.

AI and the protein corona

Implementation of standardized protocols on methodologies and analyses of protein corona can generate extensive multi-omic-based data sets that can be used to teach AI to prognosticate diseases using protein corona fingerprints and to predict protein corona formation on distinct NPs for the fundamental design of nanomedicines. Characterization and prediction of the protein corona are both important to understand the interactions of NPs in biological milieus, yet there is a large discrepancy between the two, as the former comprises substantially more reports than the latter^{124,125}. This immense difference is attributed to the disadvantages of existing high-throughput techniques and instruments used to test the biocompatibility and biofouling of nanotechnologies. Protocols for protein corona extraction can vary between laboratories, and thus there is a need for robust and standardized methods^{109,126}. Moreover, MS is laborious, expensive and requires a high level of expertise, and such high-throughput analytical experiments are subject to multiple errors and variabilities arising from laboratory-to-laboratory differences in sample preparation and analysis¹²³. Al and machine learning approaches can overcome these technical barriers and further elucidate the impact of the protein corona by predicting protein adsorption to NPs as well as their biological impacts (Box 1).

Among the promising machine learning algorithms, random forest classification (RFC), which integrates multiple decision trees to form a predictive model¹²⁷, can easily be trained with MS data sets for the identification of protein features that promote or discourage protein adsorptivity to NPs. Typical performance metrics that validate the predictions of RFCs are accuracy, precision and recall. In general, these metrics measure the overall correctness of RFC model predictions, as well as the proportion of correctly predicted positives made by the model. The RFC algorithm has been successful in predicting protein adsorption to single-walled CNTs using 38 protein features, which are variables that define the relationship between input and output data for the model, based exclusively on amino acid sequences, with 78% accuracy and 70% precision⁴⁰. Moreover, using kernel density estimates, statistical methods to estimate the distribution of variables. protein features such as elevated solvent-exposed glycine residues and high leucine residues were strongly correlated with predicted proteins that adsorb or desorb from single-walled CNTs, respectively⁴⁰. RFC has also predicted the protein corona that adsorbs on silver NPs with 75% accuracy and 76% precision¹²⁸ by predominantly using the protein physiochemical properties as features; 10 features classified protein properties, and 4 described solvent characteristics (cysteine and NaCl concentrations) and NP properties (size and zeta potential). RFCs can calculate the importance of features on the basis of how they impact model performance metrics. By excluding particular features from data sets, accuracy, precision and recall can decrease, revealing which features are necessary to have high metric values. Protein features were more important in predicting protein corona formation, but this may have been a result of the lower quantity of solvent and nanomaterial

Box 1

Machine learning contributions to NP protein corona research

Machine learning models have been developed to predict protein corona compositions and the biological impacts induced by the protein corona utilizing the following descriptors:

- Protein sequences with high affinity to nanoparticle (NP) surfaces⁴⁰
- Limited NP physicochemical properties to predict protein adsorption¹²⁸
- NP-protein interactions labelled with fluorescamine to predict protein corona¹²⁹
- Functional proteins for cellular association¹³⁰
- Protein corona to predict cellular uptake^{131,132}
- Time-dependent coronas to predict in vivo fate¹³³

features used in the model¹²⁸. Thus, more features are necessary to comparably determine those that strongly impact protein corona prediction. The majority of RFCs that predict corona formation utilize several protein-based features and limited NP descriptors. This is due to the diversity and complexity of proteins, resulting in multiple characterizable protein features (molecular weight, isoelectric point, pH, grand average of hydropathy and amino acid compositions); meanwhile, NPs have less characterizable properties that can serve as features for machine learning models. Protein corona fluorescamine labelling was used as an NP descriptor because of its positive correlation with the physiochemical properties of the NP¹²⁹. Alongside this singular NP descriptor, derived from the screening of the fluorescamine-labelled protein coronas formed on 22 diverse NPs, 4 protein classifiers were used in the RFC model for protein corona prediction with 84% precision; protein corona fluorescamine labelling serving as an NP feature was as effective as solely using typical NP features (size and charge) to build the RFC¹²⁹. Moreover, this unique NP feature was used to build a model that was able to predict the adsorption of proteins on five different 2D nanomaterials with 75% precision, a feat that has been difficult to achieve with common classifiers because of the heterogeneity of 2D nanomaterials¹²⁹. Additionally, RFCs have been used to predict the protein corona on several NPs with multiple types of surface chemistries, using proteomic data sets derived from 56 individual studies; the majority of protein coronas were predicted with >75% accuracy¹³⁰. The most important factors that dominated protein corona prediction in these models were NP surface chemistries (bare, PEGylated, functionalization of amines and other ligands). The RFC model was also used to predict protein functional components (that is, apolipoprotein, complement protein, coagulation protein, immune protein and clusterin), which were then correlated with recognition indexes impacted by the protein corona. The recognition indexes represented cell uptake efficiencies, pro-inflammatory responses and perturbations to the immune system. These findings suggest that the predicted functional components of the protein corona were associated with cell recognition of different NPs¹³⁰.

The biological impacts of protein corona-coated NPs have also been predicted using other machine learning algorithms such as partial least-squares regression (PLSR), support vector machine (SVM)-based classification, neural network (NN) and k nearest neighbour-based regression. Instead of predicting the composition of the protein corona, these algorithms use the physicochemical properties of the NP and/or the protein corona as features to predict NP-induced biological effects. Quantitative structure-activity relationship (QSAR), which is computational modelling that can correlate NP and/or protein corona physicochemical properties to biological responses, developed with PLSR predicted in vitro cell interactions of Au and Ag NPs using the protein corona as a descriptor¹³¹. Briefly, proteins identified on the coronas of a library of 105 surface-modified Au NPs were used to build single-parameter linear models describing NP-cell associations as a function of the relative abundance of each protein in the corona or as the sum of the densities of the proteins that composed the corona. Using a selection of 64 predictive proteins, PLSR predicted NP-cell associations with an accuracy of 81%; the model accuracy increased by 5% when the parameters describing the NP formulation included both protein corona and NP physiochemical parameters, suggesting that the most relevant NP parameter for the prediction of cell association was the protein corona. Moreover, the Au NP protein corona was used to build a model that predicted cell associations by Ag NPs but was not successful (accuracy ~5%). An additional model was built using only the Ag NP protein corona and resulted in a high prediction accuracy (~80%). These findings suggested that there are NP-dependent differences in protein orientation and conformation that can influence cellular associations. By calculating the similarity of the protein coronas formed around Au and Ag NPs, the core material of the NPs had the greatest influence on protein corona composition when compared with size or surface chemistry¹³¹. Compared with the aforementioned study that utilized RFC to calculate the most informative feature among both protein and NP properties¹²⁸, protein corona similarities were calculated to determine how influential the physicochemical properties of NPs were for protein corona formation¹³¹. Thus, the latter did not deduce if the protein corona was an essential feature for their cell association model. In a follow-up study, a nonlinear QSAR model was built with SVM-based classification to predict in vitro cell association based on both the physiochemical properties of 84 gold NPs, with varying surface chemistries, and the protein corona¹³². Succinctly, the nonlinear QSAR model used 6 proteins, identified from the protein coronas of 105 Au NPs, and zeta potentials as the most important contributors for predicting NP-cell associations. By including the zeta potentials of NPs, the predictive power of the NP-cell association model increased by 5%, when compared with a model that only used proteins from the corona as features (~85% accuracy)¹³². The in vivo fate of NPs was predicted with a supervised deep NN using protein coronas extracted at multiple time points from blood after in vivo circulation with 94% accuracy¹³³. The NN algorithm was taught protein corona patterns, which fluctuate over time in circulation and are dependent on NP size, to accurately predict the clearance of NPs through the spleen or liver. Moreover, the model found that NP accumulation into these organs was independent of a single protein, but rather was contingent on an assortment of proteins on the corona that formed unique patterns throughout circulation time. An adaptive lasso-identified subset of proteins, which were strongly associated with highly accurate biodistribution, could predict clearance with an analogous accuracy as the NN prediction using more than 700 proteins¹³³. SVM-based classification and k nearest neighbour-based regression algorithms have constructed quantitative nanostructure-activity relationship models of NPs to predict their biological impacts¹³⁴, but these models lack input from the protein corona. Immune activation by spherical nucleic acid nanomedicines has also been predicted with a $quantitative\,nano structure-activity\,relationship\,model^{135}, but\,similarly$ did not integrate the protein corona, which can potentially interfere with the biodistribution of the spherical nucleic acids or impact their intended therapies.

In silico approaches, such as machine learning algorithms, can effectively predict protein corona formation and biological interactions of several NPs for the tactical translation of innovative nanomedicines. By implementing predictive models to learn proteomic and other-omics data sets associated with NP protein corona (Fig. 3), we can identify protein features, such as functional protein motifs and epitopes, that may contribute to protein binding to NP surfaces as well as understand the relationship of the protein corona in NP cell recognition, protein-protein interactions and subsequent in vivo biological interactions. These extrapolative tools can also potentially predict protein binding even in the absence of new proteomic data sets. By predicting the protein corona on NPs, we can engineer nanomedicines that seamlessly integrate into the rapeutic applications without the need for experimental MS analysis. Additionally, machine learning can reduce the number of animals and experimental efforts required to assess nanomedicine efficacy before human trials.

Considering that current studies have confirmed the dependence of the protein corona on the prediction of NP-biological interactions^{131,133}, machine learning algorithms have also successfully utilized protein corona data sets to predict Alzheimer disease53 and various types of cancers^{19,26} and can potentially predict other biological processes, such as subcellular localization of NPs or the induction of epigenetic mechanisms by nanomedicines. Future efforts that support the prediction of the protein corona, as well as the comprehension of the relationships between NP physicochemical properties and corona formation, should rely on extensive libraries of well-characterized NPs and proteomic data sets of NP coronas to establish an accurate, reliable and easily accessible bioinformatic database from which data can be extracted for machine learning applications. AlphaFold, a protein structure database developed by DeepMind and EMBL-EBI, can predict protein 3D structures from amino sequences with high accuracies^{136,137} and could potentially be extended to improve the predictions of protein structures likely to form the NP protein corona and eco-corona. Furthermore, aforementioned studies emphasize the need for continued data collection to feed into machine learning models, together with developing more sophisticated models with greater predictive power, for the fabrication of safe, sustainable and effective nanomedicines.

Emerging opportunities offered by the protein corona

Although the formation of protein corona on the surface of NPs usually causes several negative consequences (such as mistargeting and inducing errors in nano-based assays), it also enables new opportunities to address a wide range of issues, from early detection of diseases to the eco-corona (Fig. 4). In other words, the recent progresses in the field of protein corona revealed the other side of the coin: the usefulness of protein corona in the design of new diagnostic and/or therapeutic techniques.

Design of new therapies

The coronavirus global pandemic underscored that a virus can modify its virulent surface protein, altering its interactions with the human immune system¹³⁸⁻¹⁴⁰. The nanomedicine community can learn from the rich literature on various interactions between biosystems and nanosized viruses, including the coronavirus, and their effects (such as immune system recognition and response). By combining these lessons with our understanding of journeys of NPs in biosystems141-144, the nanomedicine community should strive to fully and mechanistically understand biosystem interactions with corona-coated NPs, from recognition to the relevant cellular processing and pathways³². Specific attention should be paid to recognition of biosystems of NPs through the patterns and organizations of their 'rare proteins' rather than their bulk compositions. In other words, achieving a deep understanding of the critical role of patterns and organizations of less-abundant proteins in the protein corona profile of NPs enables the precise manipulation of biosystem responses to NPs and the design and development of novel and efficient protein corona therapies.

The sex and age of the biosystems are important in their responses to NPs, yet poorly considered in nanomedicine¹⁴⁵. As such, recent findings on the interactions of the coronavirus with the immune system revealed the critical roles of age¹⁴⁶ and sex¹⁴⁷ on immune responses. For example, it was shown that male-derived immune cells produce higher levels of innate immune cytokines in blood plasma compared with female-derived cells, and more robust T cell activation occurs



Fig. 3 | **Representative workflow used to predict protein corona formation with machine learning.** Protein corona data, usually from mass spectrometry experiments, are used to train a machine learning classifier that learns which features of proteins are likely to be found in versus out of the nanoparticle protein corona. Classifiers can be tested through experimental validation of singleprotein binding affinities, or by predicting and validating protein adsorption from biofluids different from those used in the training set.

in female-derived cells compared with male-derived cells during coronavirus infection.

Once the aforementioned knowledge and understanding is achieved, researchers will be more capable of designing and developing new or adapted nanotherapeutics (such as nanoimmunotherapy by activating desired immune system pathways and mRNA and gene editing nanocarriers), by precisely designing and controlling the protein corona composition and decoration of the surface of NPs. The effects of the contributing factors (such as sex and age) on the safety and efficacy of NPs are strongly dependent on the type of NPs and their potential payload (such as mRNA, proteins and active biomolecules and pharmaceuticals). For monitoring the role of sex and age on the interactions of payload-free NPs with biosystems, for example, researchers can use 'empty' coronavirus-like particles and/or membranes.



More in-depth information and analysis of the biological nanoscale recognition mechanisms and responses are offered in another perspective³². If the role of sex and age is considered in the design and development of new nanotherapeutics (based on the effects of their potential payloads), NPs with engineered biological identities can target and/or activate sex-associated and/or ageing-associated pathways, which could improve the safety and therapeutic efficacy of nanomedicine products for both sexes and all ages.

Proteomics

One of the positive attributes of the NP protein corona is that the abundance of its proteins is different from the protein composition of the native biofluid^{8,148,149}. In other words, NPs can enrich or deplete specific proteins in their corona profiles, regardless of the composition of these proteins in biological fluids, which can be useful for protein identification and characterization purposes. As such, the protein corona has a unique potential to overcome major problems in the global discovery of plasma proteomics (Box 2), such as biomarker discovery^{8,26,27,33,71,148}. The composition of protein coronas on the surface of identical NPs strongly depends on the type of disease(s) the plasma donors have (known as 'personalized' or 'disease-specific' protein corona)²⁷. This concept has been validated and used by various groups for studying personalized and disease-specific protein corona interactions with biosystems^{53,78,150-156}. Although the health condition of plasma donors can alter the protein corona profile of identical NPs²⁷, fewer proteins are identified in the corona than in the native plasma biofluid (from few tens to several hundreds in the protein corona, which depends on the physicochemical properties of NPs and the sensitivity of MS)^{8,26,123}. The low numbers of plasma proteins in protein corona relative to the native plasma biofluid may increase the sensitivity and specificity of disease and/or biomarker detection using the protein corona approach, assuming that target proteins are among the ones adsorbing to and forming the NP protein corona. For example, blood-circulating liposomes in tumour-bearing mice could capture secreted tumour-specific proteins of human cells that had been used for tumour creation³⁴.

To improve the specificity and sensitivity of disease detection, increasing the number of uniquely identified plasma proteins can be done using a 'protein corona sensor array'³³. The protein corona

sensor array combines the NP protein corona with sensor array technology and machine learning for the robust identification of biomarker patterns for the detection of diseases³³. The protein corona on the surface of various NPs (which are the sensor array elements), formed after interactions with healthy and various disease plasmas (or other biological fluids), is analysed by machine learning to identify protein/ biomolecular patterns that have critical roles in the identification and discrimination of individual diseases. The cross-reactive interactions of the protein classes with NPs may provide unique fingerprints (that is, sensor-specific biomarkers) for each type of disease, which would facilitate disease identification and discrimination³³. It is noteworthy that increasing the number of sensor array elements (by adding distinct NPs) can provide more proteomics data from plasma proteins, which, in turn, can increase the sensitivity and specificity of the machine learning algorithm for the detection, discrimination and prediction of diseases³³. The sensor array mimics the human olfactory system, which can identify and discriminate $\geq 10,000$ different odourants¹⁵⁷. However, there are nowhere near 10,000 specific receptors for the lock-and-key identification of each odourant. Instead, recognition specificity comes from pattern recognition (cross-responsive receptors that produce composite responses unique to each odourant)¹⁵⁸. Theoretically, the purpose of a sensor array is to identify, discriminate and guantify analytes and biomolecules much more sensitively and easily than specific individual sensors can. For example, there is no singular sensor capable of distinguishing NPs of different shapes and sizes. However, complex identification and discrimination of various NPs, even at very low concentration (100 ng ml⁻¹), is easily achievable by the use of a sensor array¹⁵⁹.

As in olfactory arrays, the specificity in protein corona sensor arrays comes from pattern recognition, in which the sensor array elements produce specific protein corona pattern unique to each disease type. Using liposomes with three distinct surface properties, a protein corona sensor array successfully identified and discriminated five distinct types of human cancers (lung, glioblastoma, meningioma, myeloma and pancreatic cancers) through the creation of unique protein corona patterns on NP surfaces that served as fingerprints for each type of cancer³³. By increasing the numbers of used NPs, one can expect more robustness in detection and discrimination capability of the platform. The robustness of the protein corona sensor array for

the identification and discrimination of cancers at very early stages was assessed using Golestan cohort plasmas that were obtained from healthy individuals but that would go on to be diagnosed with lung, pancreas and brain cancers several years after initial plasma collection. The outcomes revealed that the protein corona sensor array could robustly identify and discriminate among the cancers years before the patient develops clinical symptoms³³.

For disease diagnostic purposes, the main advantage of using sensor arrays is that they can recognize patterns of rare proteins rather than conventional biomarkers. This is mainly because biomarkers mostly refer to specific biomolecules (such as proteins) that are elevated rather than emergent - in the blood plasma of patients (that is, 'turning on and off' in disease and healthy conditions, respectively), each of them having a diagnostic value in the analysis of human plasma. However, the presence of proteins in the protein corona that in general are recognized as biomarkers (with high confidence being placed on protein corona purity) does not represent their elevation in blood plasma. In other words, there is less correlation between the concentration of proteins in plasma and at the surface of the NPs (that is, elevation of one protein in plasma does not cause elevation of that particular protein in the corona composition). In addition, elevation of biomarkers in plasma may substantially change the entire composition of the protein corona. Human plasma also undergoes substantial changes in the composition of small biomolecules such as metabolomes (such as glucose) and lipids (such as cholesterol)^{160,161} during disease occurrence and progress, which further alters the interaction of plasma proteins with NPs (as shown by both simulation¹⁶² and experimental¹⁶³ results). In contrast to biomarkers, the pattern recognition of rare proteins in the corona layer, associated with disease occurrence and progresses, can provide more robust diagnostic outcomes. In addition, other biomolecules in plasma (such as lipids, metabolomes and nucleic acids) can affect the patterns of rare proteins. Overall, for disease detection purposes, simply searching and finding known disease-specific biomarkers that are elevated in plasmas of patients in the protein corona profiles, rather than defining the corona pattern of rare proteins, may induce substantial errors in the sensitivity and specificity of disease identification and prediction.

In another study using silica-coated multicore superparamagnetic iron oxide NPs with various physicochemical properties, protein corona compositions were prepared, collected and subjected to mass spectrometry analysis in an automated manner for efficient proteomic profiling; over 2,000 proteins were identified from plasma samples²⁶. Because of its automated nature, the technology, which is now commercially available, could be used for the rapid and high-throughput profiling of plasma proteomes for biomarker discovery²⁶.

The unique emerging role of the NP protein corona in proteomics may be leveraged for the rapid and early screening and discrimination of various diseases (such as cancers and neurodegenerative disorders), in which very early detection could result in the early initiation of therapeutic options, improving patient quality of life and patient outcomes.

The eco-corona

Although most studies on protein corona relate to their use in nanomedicine through preclinical applications, emerging research points to the environmental aspect of the protein corona, termed the 'eco-corona' and its role in nano-ecotoxicology³⁵. The increasing production and use of NPs, including nanoplastics and nanomedicines, result in uncontrolled nanomaterial release into the environment^{164–166}. Moreover, the integration of NPs to enhance agricultural production¹⁶⁷⁻¹⁷⁰ may be another source of NP release into the environment. To fully understand how NPs biodistribute and accumulate in our environment, understanding eco-corona formation on NP surfaces is imperative. Analogous to the formation of the protein corona from blood-based biofluids in nanomedicine, the formation of the eco-corona on NPs occurs through the adsorption of proteins¹⁷¹, humic substances^{23,172-175}, metabolites¹⁷¹ and natural organic matter (NOM)¹⁷⁶. The formation of the eco-corona outside aquatic organisms, as well as within organisms following NP internalization, was reviewed elsewhere³⁵. Biofluid-derived protein corona studies, which have studied the interactions between nanomaterials and biomolecules in complex biological fluids, can be instrumental for analysing and understanding the real-world ecocorona. Moreover, extracting the real-world eco-corona is challenging and complex, as NPs transgress among industrial, aquatic, terrestrial and atmospheric environments. Box 3 features and relates eco-corona and biofluid-derived protein corona research.

Characterization of adsorbed amphiphiles (that is, humic substances, peptides, fatty acids and NOM) on the eco-corona is deficient because of the extensive heterogeneity of amphiphiles in the environment. It was found that amphiphiles derived from algal exudates exhibit Vroman-like competitive binding, in which proteins with high binding affinities displace proteins with low binding affinities, on nanosheet surfaces¹²⁷. Soft and hard eco-corona form with amphiphiles possessing varying binding affinities to the graphene nanosheets. The protein component on the eco-corona is frequently scrutinized as proteins can engage with receptors and influence biological signalling

Box 2

Proteomics shortcomings for biomarker discovery

Biomolecular analysis of blood plasma (such as electrolytes, small molecules, nucleic acids, drugs and proteins) is the most widespread diagnostic procedure in medicine. Although the composition of circulating plasma biomolecules is dynamic across the spectrum of health and disease, there is a trade-off between depth of coverage and sample throughput, given the vast dynamic range and high complexity of the plasma proteome²⁰⁰⁻²⁰². One of the main issues with finding a robust plasma biomarker is the existence of abundant proteins (such as albumin that makes up roughly half of the total protein mass, apolipoproteins, acute phase proteins and proteins of the coagulation cascade) that can substantially reduce the sensitivity of mass spectrometry approaches towards the identification of the variation of very low-abundance proteins that can be considered biomarkers (which are at the range of picogram per millilitre, approximately 10¹⁰-fold lower plasma dynamic range compared with the concentration of albumin, which is ~35–50 mg ml⁻¹)^{203,204}. Various strategies (such as depleting high-abundance proteins) are being used to overcome this major proteomics issue in the discovery of new plasma biomarkers, which further improves the depth of proteome coverage and the robustness of outcomes^{200,205-207}.

Box 3

Eco-corona versus biofluid protein corona

Similarities

Analogous to the interactions of nanoparticles (NPs) in biological matrices forming the protein corona^{19,57,59}, NPs introduced into environmental fluids can interact with ecological biomolecules resulting in the formation of the eco-corona³⁵⁻³⁷. Specifically, NP physicochemical properties influence formation of both the biofluid protein corona⁵⁷ and the composition of the eco-corona²⁰⁸. Likewise, the conditions of organism-derived or environmental-derived matrices can impact the corona profile^{209,210}. Moreover, NP-biological interactions are highly complex regardless of how the corona was produced; intracellular exchange of NP protein corona can disturb proteostasis in HeLa cells²¹¹; release of cellular proteins into the extracellular matrix can induce the exchange of NP protein corona components and alter cell uptake²¹²; and toxicity has been reported in Daphnia magna because of the 'Trojan horse-like' influence of the eco-corona prompting increases of zinc oxide NP uptake and dissolution²¹³. Eco-corona adsorption increased NP consumption by D. magna, and the acidic conditions in the guts of D. magna markedly increased NP dissolution and toxicity and thus a 'Trojan horse' effect was observed.

Differences

Apart from proteins, both the eco-corona and the biofluid corona can be composed of an assortment of constituents. However, the

in organisms and may impact ecological systems through biotransformation and environmental distribution^{35,178}. Comparably to the biofluid-derived protein corona detailed earlier, adsorbed ecological substances on NP surfaces can alter identity, properties and biological impacts of NPs^{179,180}. For example, secreted ecological proteins from planktonic Daphnia magna formed an eco-corona on polystyrene NPs, which then become more toxic than their pristine counterparts in vivo; the clearance and ability of D. magna to feed were negatively impacted following exposure to the eco-corona-coated plastic NPs¹⁷¹. Adsorption of humic acid on zinc oxide NPs increased the hatching success of NPexposed zebrafish embryos. However, NP toxicity was not ameliorated by the eco-corona¹⁷². Attenuated NP toxicity towards *D. magna* was observed following exposure to eco-corona-coated tungsten carbide cobalt NPs; the reduced toxicity was speculated to be mediated through NP-organism interactions after the NOM-rich eco-corona NPs were internalized¹⁸¹. Similarly, formation of the eco-corona on silver NPs, through the adsorption of NOM collected from river water, decreased the bactericidal efficacy of NPs in the bacteria Shewanella oneidensis MR-1 (ref. 182). It is of vital importance to characterize NP eco-coronas to establish the environmental fate and eco-toxicity of NPs and engineer safer nanotechnologies, such as innocuous agricultural sensors with enhanced efficacy^{183,184}. Characterizations of the protein corona in human biofluids have enabled NPs to gain much traction in clinical practice, yet the eco-corona remains much more sparsely characterized within a much larger exposure space. As such, the current state of the field supports the need to develop standardized protocols and

eco-corona incorporates heterogeneous environmental components that are devoid in the biofluid corona, such as humic substances (HS) (that is, natural organic matter, fluvic and humic acids)²³ and polymeric molecules secreted as metabolic by-products by aquatic organisms (bacterial gut secretions, undigested organic matter, kairomones, polysaccharides and metabolites expelled by D. magna and other aquatic species)^{171,214}. Furthermore, there exist substantial chemical and structural differences between soil and aquatic HS¹⁷⁴ that can hypothetically result in different coronas. Likewise, secreted by-products from other aquatic and microbial species can differ drastically, adding to the complexity of the eco-corona^{36,179,215}. Although the protein corona derived from biological fluids has been abundantly studied relative to the eco-corona, there is much more to be explored in the latter to comprehend the implications of these complexes in ecosystems. As NPs enter the environment, through agricultural processes, waste disposal runoff and industrial wastewater discharge, the eco-corona composition is likely to fluctuate. For example, as mentioned earlier, D. magna can ingest NPs coated with eco-coronas. It is possible that these ingested NPs can subsequently enter the food chain, accumulate in aquatic organisms and potentially be transferred to humans through the consumption of these organisms such as fish. The implications of the eco-corona on human health are not well understood and require extensive study.

improved technologies to extract and assess the eco-corona and its biological and ecological impacts under environmentally realistic conditions, a phenomenon that has not been feasible to date (Box 3).

It is now well understood that the environmental release of engineered NPs, whether intentional (such as in the application of NPs for environmental or agricultural applications) or unintentional (such as through product degradation), is inevitable^{35,164,185}. NPs interact with a wide range of environmental components, from air and water to live organisms including bacteria and plants, in a component-specific manner. In addition, NPs can easily travel among various environmental components and even get into the food web. For example, NPs in air, soil and/or water can easily transfer into plants, vegetables, fruits and fish and interact with them on various levels on the basis of the organism, plant, vegetable or fruit type^{164,186-189}. Ingestion of environmentally relevant nanoplastics^{190,191} can increase fat absorption by gastrointestinal cells modulated by the biomolecular corona of digested nanoplastics³⁹. Therefore, from an environmental and food safety, monitoring and management perspective, it is essential to understand the environmental journey of NPs.

A deeper understanding of the formation of the eco-corona at the surface of NPs may provide essential information regarding their environmental transfer before and after their uptake by an organism and even their translocation between various organisms or species. Before entering an environmental milieu, the eco-corona may consist of a mixture of (bio)molecules originating from previous exposure to an organism, whereby the eco-corona will accumulate various types of

biomolecules including proteins, lipids and metabolomes, depending to the organism type¹⁹². For example, in plants, the apoplast that comprises the intercellular space, the cell walls and the xylem, a vascular tissue, contains various types of proteins, lipids and other types of plant-specific biomolecules¹⁹². Therefore, the analysis of eco-corona of NPs may shed light on environmental journey of NPs and can assist decision makers on environmental and food safety, monitoring and management, enabling them to take proper actions regarding the current challenges in the field of environmental nanotoxicity.

Conclusion and perspectives

The mechanistic understanding of the protein corona, which has been the object of the majority of biomolecular corona studies thus far, has enabled the development of more efficient and safe nanotechnologies that can bridge nano-technological advances and clinical applications. Achieving robust characterization of the formation and evolution of the protein corona in various environments not only improves our understanding of the interactions between biosystems and NPs but also enables the prediction of such interactions with high accuracy. Although protein corona formation creates inertia in clinical translation of nanomedicinal technologies, the protein corona also creates new opportunities for multiplexed diagnostics and for protein corona engineering for targeted delivery and sensing applications. The dynamic nature of the protein corona also provides a paradigm shift in our understanding of the unintended environmental journey of NPs. The adsorption of environmental proteins and natural organic matter onto nanomaterials intentionally put into practice for agricultural applications, or indirectly introduced into the environment via manufacturing processes and waste disposal, is not stagnant; the eco-corona can undergo various transformations and ecological interactions as nanomaterials transit the environment.

Achieving reproducible data sets across research groups involved in protein corona research remains one of the major issues in the field. This poor reproducibility is, at least in large part, due to the variability of biological systems from which the protein corona has been studied (such as, for blood plasma, sex, gender, age, ethnicity and health spectrum)^{99,145}, the lack of unifying standards for nano-bio characterization protocols¹⁹³ (specifically for MS)¹²³ and variabilities in experimental reporting strategies^{194,195}. The nanomedicine community would also benefit from the development of new methodologies to probe the conformation of proteins in the biomolecular corona layer. Although some techniques, such as circular dichroism, can probe the conformational changes of single proteins after interactions with the surface of NPs, robust methods to probe conformational changes of various proteins in the corona layer are lacking. The information on the multiprotein conformational changes in the biomolecular corona layer helps the understanding and prediction of recognition of NPs by immune systems and their responses to nanomedicine technologies.

Another issue in protein corona research is that less attention has been paid to the role of the patterns and organizations of lowabundance proteins and biomolecules, when compared with the bulk corona compositions, in their interactions with biosystems. Robust and precise characterization can lead to a deep understanding of biosystem responses enabling the development of new, safe and efficient therapeutic approaches. In addition, the critical role of disease-specific lipids (such as cholesterol)¹⁶² and metabolomes (such as glucose)¹⁶², which can substantially alter the interaction of plasma proteins with NPs and, consequently, affect the patterns and organizations of the rare proteins and biomolecules in the corona layer, should be studied and considered. A further obstacle is the lack of thorough investigation of protein corona profiles of certain NPs that are widely used in clinical trials or clinics. For example, achieving robust protein corona characterization on lipid NPs (specifically those used for COVID-19 vaccines)¹⁹⁶ and NPs that are being used for payload delivery to the central nervous system¹⁹⁷ will enable researchers to better understand and predict biosystem responses to the NPs and, therefore, develop safer and more efficient nanotherapeutics.

Although the investigation of NPs in human biofluids has been an ongoing field of research over the past few decades, the use of nanotechnologies in plants and agriculture is relatively nascent¹⁸³. Environmental stressors, such as climate change, pathogens and population growth, complexify agricultural practices. Nanobiotechnology offers sustainable solutions by adapting nanomaterials as nanosensors that can monitor plant analytes, augment crop stress resistance and proctor productivity by reporting metrics that can advance agronomy. Nanomaterials have also served as carriers of plasmid DNA and small interfering RNA¹⁶⁸ for genetic and post-transcriptional biofortification of crop plants. NPs have contributed to pesticide and nutrient delivery¹⁹⁸ and as elicitors of plant-defence responses¹⁹⁹. Despite the prolific and growing use of nanotechnology in agriculture, the nanobio interactions in nanotechnologies remain unknown. In plants, the use of nanotechnologies results in spontaneous biomolecule adsorption from plant fluids onto nanosensor surfaces. The protein corona that forms on NPs in plants remains largely unexplored yet dictates NP interactions with plants and the environment¹⁸³. The growing interest for nanotechnologies in agriculture motivates further exploration of the fundamental nano-bio interactions that drive spontaneous protein and eco-corona formation before deploying these nanotechnologies into the field.

Regardless of the NP application space, whether in the clinic or in the environment, experimental approaches and biological variability in characterizing the NP protein corona across different systems limit the throughput of experimental validation of nanotechnologies. The protein corona has vast implications when applying NPs in biological systems, unpredictably changing outcomes such as the NP function, biodistribution and toxicity. Yet, to-date, there are orders of magnitude more reports of nanotechnologies developed for supposed biological application than reports of their interactions with biologically relevant milieus. This discrepancy is seen because testing nanotechnologies for biocompatibility and biofouling is a costly and time-consuming process that often relies on high-throughput MS or similar experiments with lower throughput. By implementing AI and machine learning algorithms, we can identify protein features that contribute to protein adsorption or omission from the NP protein corona^{128,130}. A well-trained machine learning classifier may also enable rapid determination of protein corona composition from entirely new biofluid or NP data sets (that is, unfamiliar relative to the training data), which can be confirmed experimentally using surface binding measurement assays⁴⁰. In this manner, machine learning supports the development of predictive protein corona models that will enable researchers to implement a wider range of nanotechnologies across different biological environments. Developments of supervised learning tools provide algorithms that researchers can use to parse protein properties from publicly available databases to determine protein corona formation, as a step towards in silico testing of nanotechnologies for their biocompatibility and biofouling propensities in a broad range of applications.

Published online: 24 March 2023

References

- Ke, P. C., Lin, S., Parak, W. J., Davis, T. P. & Caruso, F. A decade of the protein corona. ACS Nano 11, 11773–11776 (2017).
- Lima, T., Bernfur, K., Vilanova, M. & Cedervall, T. Understanding the lipid and protein corona formation on different sized polymeric nanoparticles. Sci. Rep. 10, 1129 (2020).
- Wan, S. et al. The 'sweet' side of the protein corona: effects of glycosylation on nanoparticle-cell interactions. ACS Nano 9, 2157–2166 (2015).
- Martel, J. et al. Fatty acids and small organic compounds bind to mineralo-organic nanoparticles derived from human body fluids as revealed by metabolomic analysis. *Nanoscale* 8, 5537–5545 (2016).
- Chetwynd, A. J., Zhang, W., Thorn, J. A., Lynch, I. & Ramautar, R. The nanomaterial metabolite corona determined using a quantitative metabolomics approach: a pilot study. Small 16, e2000295 (2020).
- Zhang, W., Chetwynd, A. J., Thorn, J. A., Lynch, I. & Ramautar, R. Understanding the significance of sample preparation in studies of the nanoparticle metabolite corona. ACS Meas. Au 2, 251–260 (2022).
- Lynch, I., Dawson, K. A. & Linse, S. Detecting cryptic epitopes created by nanoparticles. Sci. STKE 2006, pe14 (2006).
- Monopoli, M. P., Åberg, C., Salvati, A. & Dawson, K. A. Biomolecular coronas provide the biological identity of nanosized materials. *Nat. Nanotechnol.* 7, 779–786 (2012).
 Walczyk, D., Bombelli, F. B., Monopoli, M., Lynch, I. & Dawson, K. A. What the 'cell' sees
- in bionanoscience. J. Am. Chem. Soc. 132, 5761–5768 (2010).
 Hadiidemetriou. M. & Kostarelos, K. Evolution of the nanoparticle corona. Nat. Nanotechnol.
- Hadjidemetriou, M. & Kostarelos, K. Evolution of the hanoparticle corona. Nat. Nanotecrim 12, 288–290 (2017).
- 11. Chetwynd, A. J. & Lynch, I. The rise of the nanomaterial metabolite corona, and emergence of the complete corona. *Environ. Sci. Nano* **7**, 1041–1060 (2020).
- Cedervall, T. et al. Understanding the nanoparticle-protein corona using methods to quantify exchange rates and affinities of proteins for nanoparticles. Proc. Natl Acad. Sci. USA 104, 2050–2055 (2007).
- Bangham, A. D., Pethica, B. A. & Seaman, G. V. The charged groups at the interface of some blood cells. *Biochem. J.* 69, 12–19 (1958).
- Vroman, L. Effect of adsorbed proteins on the wettability of hydrophilic and hydrophobic solids. *Nature* 196, 476–477 (1962).
- Harley, J. D. & Margolis, J. Haemolytic activity of colloidal silica. Nature 189, 1010–1011 (1961).
- Zhang, X. et al. Intracellular activation of bioorthogonal nanozymes through endosomal proteolysis of the protein corona. ACS Nano 14, 4767–4773 (2020).
- 17. Francia, V. et al. Corona composition can affect the mechanisms cells use to internalize nanoparticles. ACS Nano 13, 11107–11121 (2019).
- Lesniak, A. et al. Effects of the presence or absence of a protein corona on silica nanoparticle uptake and impact on cells. ACS Nano 6, 5845–5857 (2012).
- Bertrand, N. et al. Mechanistic understanding of in vivo protein corona formation on polymeric nanoparticles and impact on pharmacokinetics. *Nat. Commun.* 8, 777 (2017).
- Salvati, A. et al. Transferrin-functionalized nanoparticles lose their targeting capabilities when a biomolecule corona adsorbs on the surface. Nat. Nanotechnol. 8, 137–143 (2013).
- Chen, F. et al. Complement proteins bind to nanoparticle protein corona and undergo dynamic exchange in vivo. Nat. Nanotechnol. 12, 387–393 (2017).
- 22. Ge, C. et al. Binding of blood proteins to carbon nanotubes reduces cytotoxicity. *Proc. Natl Acad. Sci. USA* **108**, 16968–16973 (2011).
- Fadare, O. O. et al. Eco-corona vs protein corona: effects of humic substances on corona formation and nanoplastic particle toxicity in *Daphnia magna*. Environ. Sci. Technol. 54, 8001–8009 (2020).
- Trinh, D. N. et al. Nanoparticle biomolecular corona-based enrichment of plasma glycoproteins for N-glycan profiling and application in biomarker discovery. ACS Nano 16, 5463–5475 (2022).
- Papafilippou, L., Claxton, A., Dark, P., Kostarelos, K. & Hadjidemetriou, M. Protein corona fingerprinting to differentiate sepsis from non-infectious systemic inflammation. *Nanoscale* 12, 10240–10253 (2020).
- Blume, J. E. et al. Rapid, deep and precise profiling of the plasma proteome with multi-nanoparticle protein corona. *Nat. Commun.* 11, 3662 (2020).
- Hajipour, M. J., Laurent, S., Aghaie, A., Rezaee, F. & Mahmoudi, M. Personalized protein coronas: a 'key' factor at the nanobiointerface. *Biomater. Sci.* 2, 1210–1221 (2014).
- Wang, X. et al. Chiral surface of nanoparticles determines the orientation of adsorbed transferrin and its interaction with receptors. ACS Nano 11, 4606–4616 (2017).
- Mirshafiee, V., Kim, R., Park, S., Mahmoudi, M. & Kraft, M. L. Impact of protein pre-coating on the protein corona composition and nanoparticle cellular uptake. *Biomaterials* 75, 295–304 (2016).
- Ritz, S. et al. Protein corona of nanoparticles: distinct proteins regulate the cellular uptake. *Biomacromolecules* 16, 1311–1321 (2015).
- Mohr, K. et al. Aggregation behavior of polystyrene-nanoparticles in human blood serum and its impact on the in vivo distribution in mice. J. Nanomed. Nanotechnol. 5, 1000193 (2014).
- Dawson, K. A. & Yan, Y. Current understanding of biological identity at the nanoscale and future prospects. Nat. Nanotechnol. 16, 229–242 (2021).

This article elaborates on how biological systems interact with nanomaterials and protein coronas and form bionanoscale synapses. The authors discuss future directions within protein corona research, such as the development of new technologies to study NP-biological interactions in cells.

- Caracciolo, G. et al. Disease-specific protein corona sensor arrays may have disease detection capacity. *Nanoscale Horiz.* 4, 1063–1076 (2019).
- Hadjidemetriou, M., Al-ahmady, Z., Buggio, M., Swift, J. & Kostarelos, K. A novel scavenging tool for cancer biomarker discovery based on the blood-circulating nanoparticle protein corona. *Biomaterials* 188, 118–129 (2019).
- Wheeler, K. E. et al. Environmental dimensions of the protein corona. *Nat. Nanotechnol.* 16, 617–629 (2021).
- Nasser, F., Constantinou, J. & Lynch, I. Nanomaterials in the environment acquire an "eco-corona" impacting their toxicity to *Daphnia magna* – a call for updating toxicity testing policies. *Proteomics* 20, e1800412 (2020).
- Lynch, I., Dawson, K. A., Lead, J. R. & Valsami-Jones, E. in Frontiers of Nanoscience Vol. 7 (eds Lead. J. R. & Valsami-Jones, E.) 127–156 (Elsevier, 2014).
- Ramsperger, A. F. R. M. et al. Environmental exposure enhances the internalization of microplastic particles into cells. Sci. Adv. 6. eabd1211 (2020).
- Deloid, G. M. et al. Incineration-generated polyethylene micro-nanoplastics increase triglyceride lipolysis and absorption in an in vitro small intestinal epithelium model. *Environ. Sci. Technol.* 56, 12288–12297 (2022).
- Ouassil, N., Pinals, R. L., Del Bonis-O'Donnell, J. T., Wang, J. W. & Landry, M. P. Supervised learning model predicts protein adsorption to carbon nanotubes. *Sci. Adv.* 8, eabm0898 (2022).

This study utilizes machine learning algorithms to predict protein corona formation on CNTs by solely utilizing protein sequences from proteomic data sets. Moreover, Al identifies the most important protein features associated with high binding affinity to CNTs.

- Blunk, T., Hochstrasser, D. F., Sanchez, J. -C., Müller, B. W. & Müller, R. H. Colloidal carriers for intravenous drug targeting: plasma protein adsorption patterns on surface-modified latex particles evaluated by two-dimensional polyacrylamide gel electrophoresis. *Electrophoresis* 14, 1382–1387 (1993).
- Norman, M. E., Williams, P. & Illum, L. In vivo evaluation of protein adsorption to sterically stabilised colloidal carriers. J. Biomed. Mater. Res. 27, 8610866 (1993).
- Norman, M. E., Williams, P. & Illum, L. Influence of block copolymers on the adsorption of plasma proteins to microspheres. *Biomaterials* 14, 193–202 (1993).
- Gebauer, J. S. et al. Impact of the nanoparticle-protein corona on colloidal stability and protein structure. *Langmuir* 28, 9673–9679 (2012).
- Pan, H., Qin, M., Meng, W., Cao, Y. & Wang, W. How do proteins unfold upon adsorption on nanoparticle surfaces? *Langmuir* 28, 12779–12787 (2012).
- Roach, P., Farrar, D. & Perry, C. C. Interpretation of protein adsorption: surface-induced conformational changes. J. Am. Chem. Soc. 127, 8168–8173 (2005).
- Roach, P., Farrar, D. & Perry, C. C. Surface tailoring for controlled protein adsorption: effect of topography at the nanometer scale and chemistry. J. Am. Chem. Soc. 128, 3939–3945 (2006).
- Ashby, J., Duan, Y., Ligans, E., Tamsi, M. & Zhong, W. High-throughput profiling of nanoparticle–protein interactions by fluorescamine labeling. *Anal. Chem.* 87, 2213–2219 (2015).
- Kelly, P. M. et al. Mapping protein binding sites on the biomolecular corona of nanoparticles. Nat. Nanotechnol. 10, 472–479 (2015).
- Castagnola, V. et al. Biological recognition of graphene nanoflakes. Nat. Commun. 9, 1577 (2018).
- Lara, S. et al. Identification of receptor binding to the biomolecular corona of nanoparticles. ACS Nano 11, 1884–1893 (2017).
- 52. Ren, J. et al. Precision nanomedicine development based on specific opsonization of human cancer patient-personalized protein coronas. *Nano Lett.* **19**, 4692–4701 (2019).
- Corbo, C. et al. Analysis of the human plasma proteome using multi-nanoparticle protein corona for detection of Alzheimer's disease. *Adv. Healthc. Mater.* 10, e2000948 (2021).
- Colapicchioni, V. et al. Personalized liposome-protein corona in the blood of breast, gastric and pancreatic cancer patients. Int. J. Biochem. Cell Biol. 75, 180–187 (2016).
- Pink, M., Verma, N., Kersch, C. & Schmitz-Spanke, S. Identification and characterization of small organic compounds within the corona formed around engineered nanoparticles. *Environ. Sci. Nano* 5, 1420–1427 (2018).
- Baig, N., Kammakakam, I., Falath, W. & Kammakakam, I. Nanomaterials: a review of synthesis methods, properties, recent progress, and challenges. *Mater. Adv.* 2, 1821–1871 (2021).
- Lundqvist, M. et al. Nanoparticle size and surface properties determine the protein corona with possible implications for biological impacts. *Proc. Natl Acad. Sci. USA* **105**, 14265–14270 (2008).
- Ashby, J., Pan, S. & Zhong, W. Size and surface functionalization of iron oxide nanoparticles influence the composition and dynamic nature of their protein corona. ACS Appl. Mater. Interfaces 6, 15412–15419 (2014).
- García-Álvarez, R., Hadjidemetriou, M., Sánchez-Iglesias, A., Liz-Marzán, L. M. & Kostarelos, K. In vivo formation of protein corona on gold nanoparticles: the effect of their size and shape. Nanoscale 10, 1256–1264 (2018).
- Monopoli, M. P. et al. Physical-chemical aspects of protein corona: relevance to in vitro and in vivo biological impacts of nanoparticles. J. Am. Chem. Soc. 133, 2525–2534 (2011).
- Gorshkov, V., Bubis, J. A., Solovyeva, E. M., Gorshkov, M. V. & Kjeldsen, F. Protein corona formed on silver nanoparticles in blood plasma is highly selective and resistant to physicochemical changes of the solution. *Environ. Sci. Nano* 6, 1089–1098 (2019).
- Maiorano, G. et al. Effects of cell culture media on the dynamic formation of proteinnanoparticle complexes and influence on the cellular response. ACS Nano 4, 7481–7491 (2010).

- Mahmoudi, M. et al. Temperature: the 'ignored' at the NanoBio interface. ACS Nano 7, 6555–6562 (2013).
- Casals, E., Pfaller, T., Duschl, A., Oostingh, G. J. & Puntes, V. Time evolution of the nanoparticle protein corona. ACS Nano 4, 3623–3632 (2010).
- 65. Abbina, S. et al. Blood circulation of soft nanomaterials is governed by dynamic remodeling of protein opsonins at nano-biointerface. *Nat. Commun.* **11**, 3048 (2020). The in vivo protein corona is analysed over time to understand mechanisms that increase the blood residency time of NPs. This study confirmes that the evolution of the in vivo protein corona is highly dynamic and the corona, composed of several opsonins, fluctuates as NPs circulate in the blood.
- Dell'Orco, D., Lundqvist, M., Oslakovic, C., Cedervall, T. & Linse, S. Modeling the time evolution of the nanoparticle-protein corona in a body fluid. *PLoS ONE* 5, e10949 (2010).
- Nel, A. E. et al. Understanding biophysicochemical interactions at the nano-bio interface. Nat. Mater. 8, 543–557 (2009).
- Lesniak, A. et al. Nanoparticle adhesion to the cell membrane and its effect on nanoparticle uptake efficiency. J. Am. Chem. Soc. 135, 1438–1444 (2013).
- Deng, Z. J., Liang, M., Monteiro, M., Toth, I. & Minchin, R. F. Nanoparticle-induced unfolding of fibrinogen promotes Mac-1 receptor activation and inflammation. *Nat. Nanotechnol.* 6, 39–44 (2011).
- Li, M. et al. Nanoparticle elasticity affects systemic circulation lifetime by modulating adsorption of apolipoprotein A-I in corona formation. Nat. Commun. 13, 4137 (2022).
 The protein corona analysis of lipid-based core-shell NPs reveales that apolipoprotein A1 correlated with blood clearance lifetimes of elastic NPs.
- Tenzer, S. et al. Rapid formation of plasma protein corona critically affects nanoparticle pathophysiology. Nat. Nanotechnol. 8, 772–781 (2013).
- 72. Carril, M. et al. In situ detection of the protein corona in complex environments. *Nat. Commun.* **8**, 1542 (2017).
- Milani, S., Baldelli Bombelli, F., Pitek, A. S., Dawson, K. A. & R\u00e4dler, J. Reversible versus irreversible binding of transferrin to polystyrene nanoparticles: soft and hard corona. ACS Nano 6, 2532–2541 (2012).
- Kokkinopoulou, M., Simon, J., Landfester, K., Mailänder, V. & Lieberwirth, I. Visualization of the protein corona: towards a biomolecular understanding of nanoparticle–cell interactions. *Nanoscale* 9, 8858–8870 (2017).
- De, M., You, C. C., Srivastava, S. & Rotello, V. M. Biomimetic interactions of proteins with functionalized nanoparticles: a thermodynamic study. J. Am. Chem. Soc. 129, 10747–10753 (2007).
- 76. Vilanova, O. et al. Understanding the kinetics of protein–nanoparticle corona formation. ACS Nano 10, 10842–10850 (2016).
- Zhang, Y., Wu, J. L. Y., Lazarovits, J. & Chan, W. C. W. An analysis of the binding function and structural organization of the protein corona. J. Am. Chem. Soc. 142, 8827–8836 (2020).
- Mohammad-Beigi, H. et al. Mapping and identification of soft corona proteins at nanoparticles and their impact on cellular association. *Nat. Commun.* **11**, 4535 (2020).
 By utilizing in situ click chemistry, the interactions between soft corona proteins and cellular interfaces are analysed. This study confirms that although soft corona proteins can also comprise the hard corona, soft corona proteins modulate NP-cell interactions owing to their dynamic nature.
- 79. Baimanov, D. et al. In situ analysis of nanoparticle soft corona and dynamic evolution. Nat. Commun. 13, 5389 (2022).
 In this study, chiral NPs differentiate soft and hard corona proteins and suggest that protein corona formation is stereo-specific. Chirality-dependent protein coronas are found to mediate different NP blood circulation lifetimes, suggesting that protein coronas evolve in vivo leading to rapid clearance from circulation.
- Liu, J., Sonshine, D. A., Shervani, S. & Hurt, R. H. Controlled release of biologically active silver from nanosilver surfaces. ACS Nano 4, 6903–6913 (2010).
- Levard, C. et al. Sulfidation of silver nanoparticles: natural antidote to their toxicity. Environ. Sci. Technol. 47, 13440–13448 (2013).
- Miclāuş, T. et al. Dynamic protein coronas revealed as a modulator of silver nanoparticle sulphidation in vitro. Nat. Commun. 7, 11770 (2016).
- Yin, H. et al. Reducing the cytotoxicity of ZnO nanoparticles by a pre-formed protein corona in a supplemented cell culture medium. RSC Adv. 5, 73963–73973 (2015).
- Duan, G. et al. Protein corona mitigates the cytotoxicity of graphene oxide by reducing its physical interaction with cell membrane. *Nanoscale* 7, 15214–15224 (2015).
- Leibe, R. et al. The protein corona suppresses the cytotoxic and pro-inflammatory response in lung epithelial cells and macrophages upon exposure to nanosilica. Arch. *Toxicol.* 93, 871–885 (2019).
- Coreas, R. et al. Biological impacts of reduced graphene oxide affected by protein corona formation. Chem. Res. Toxicol. 35, 1244–1256 (2022).
- Barbalinardo, M., Caicci, F., Cavallini, M. & Gentili, D. Protein corona mediated uptake and cytotoxicity of silver nanoparticles in mouse embryonic fibroblast. Small 14, 1801219 (2018).
- Li, Y. et al. Protein corona of airborne nanoscale PM2.5 induces aberrant proliferation of human lung fibroblasts based on a 3D organotypic culture. *Sci. Rep.* 8, 1939 (2018).
 Liu, N., Tang, M. & Ding, J. The interaction between nanoparticles-protein corona
- De Jack, W. & Bing, S. The inclusion occurs in an operation of the second second
- nanotubes with human blood platelets. *Biomaterials* **35**, 6182–6194 (2014). 91. Yan, Y. et al. Differential roles of the protein corona in the cellular uptake of nanoporous
- Yan, Y. et al. Differential roles of the protein corona in the cetular uptake of nanoporous polymer particles by monocyte and macrophage cell lines. ACS Nano 7, 10960–10970 (2013).

- 92. Park, J. Y. et al. Unfolded protein corona surrounding nanotubes influence the innate and adaptive immune system. Adv. Sci. 8, 2004979 (2021).
- Latreille, P. L. et al. Scratching the surface of the protein corona: challenging measurements and controversies. ACS Nano 16, 1689–1707 (2022).
- Moein Moghimi, S., Simberg, D., Skotland, T., Yaghmur, A. & Christy Hunter, A. The interplay between blood proteins, complement, and macrophages on nanomedicine performance and responses. J. Pharmacol. Exp. Ther. 370, 581–592 (2019).
- Moghimi, S. M., Simberg, D., Papini, E. & Farhangrazi, Z. S. Complement activation by drug carriers and particulate pharmaceuticals: principles, challenges and opportunities. *Adv. Drug Deliv. Rev.* 157, 83–95 (2020).
- Ashkarran, A. A., Dararatana, N., Crespy, D., Caracciolo, G. & Mahmoudi, M. Mapping the heterogeneity of protein corona by ex vivo magnetic levitation. *Nanoscale* 12, 2374–2383 (2020).
- 97. Forest, V. & Pourchez, J. The nanoparticle protein corona: the myth of average. Nano Today **11**, 700–703 (2016).
- Levsky, J. Gene expression and the myth of the average cell. Trends Cell Biol. 13, 4–6 (2003).
- Leong, H. S. et al. On the issue of transparency and reproducibility in nanomedicine. Nat. Nanotechnol. 14, 629–635 (2019).
- Moyano, D. F. et al. Fabrication of corona-free nanoparticles with tunable hydrophobicity. ACS Nano 8, 6748–6755 (2014).
- Safavi-Sohi, R. et al. Bypassing protein corona issue on active targeting: zwitterionic coatings dictate specific interactions of targeting moieties and cell receptors. ACS Appl. Mater. Interfaces 8, 22808–22818 (2016).
- Pattipeiluhu, R. et al. Unbiased identification of the liposome protein corona using photoaffinity-based chemoproteomics. ACS Cent. Sci. 6, 535–545 (2020).
- Walkey, C. D., Olsen, J. B., Guo, H., Emili, A. & Chan, W. C. W. Nanoparticle size and surface chemistry determine serum protein adsorption and macrophage uptake. J. Am. Chem. Soc. 134, 2139–2147 (2012).
- Schöttler, S. et al. Protein adsorption is required for stealth effect of poly(ethylene glycol)- and poly(phosphoester)-coated nanocarriers. *Nat. Nanotechnol.* 11, 372–377 (2016).
- Torchilin, V. P., Berdichevsky, V. R., Barsukov, A. A. & Smirnov, V. N. Coating liposomes with protein decreases their capture by macrophages. *FEBS Lett.* **111**, 184–1888 (1980).
- Zhang, Z. et al. Brain-targeted drug delivery by manipulating protein corona functions. Nat. Commun. 10, 3561 (2019).
- Tonigold, M. et al. Pre-adsorption of antibodies enables targeting of nanocarriers despite a biomolecular corona. Nat. Nanotechnol. 13, 862–869 (2018).
- Stordy, B. et al. Conjugating ligands to an equilibrated nanoparticle protein corona enables cell targeting in serum. *Chem. Mater*, **34**, 6868–6882 (2022).
- Mahmoudi, M. The need for improved methodology in protein corona analysis. Nat. Commun. 13, 49 (2022).

This comment remarks on the need to improve methodologies for protein corona characterization that enhance reproducibility and transparency in nanomedicine, outlines common sources of errors within methodology and data analysis and elaborates on how methodological errors can be reduced.

- Mahmoudi, M. et al. A new approach for the in vitro identification of the cytotoxicity of superparamagnetic iron oxide nanoparticles. *Colloids Surf. B Biointerfaces* 75, 300–309 (2010).
- Cabaleiro-Lago, C. et al. Inhibition of amyloid β protein fibrillation by polymeric nanoparticles. J. Am. Chem. Soc. 130, 15437–15443 (2008).
- Linse, S. et al. Nucleation of protein fibrillation by nanoparticles. Proc. Natl Acad. Sci. USA 104, 8691–8696 (2007).
- McClain, S. M., Ojoawo, A. M., Lin, W., Rienstra, C. M. & Murphy, C. J. Interaction of alphasynuclein and its mutants with rigid lipid vesicle mimics of varying surface curvature. ACS Nano 14, 10153–10167 (2020).
- Mahmoudi, M. et al. The protein corona mediates the impact of nanomaterials and slows amyloid beta fibrillation. ChemBioChem 14, 568–572 (2013).
- Li, H. et al. The protein corona and its effects on nanoparticle-based drug delivery systems. Acta Biomater. 129, 57–72 (2021).
- Kah, J. C. Y., Chen, J., Zubieta, A. & Hamad-Schifferli, K. Exploiting the protein corona around gold nanorods for loading and triggered release. ACS Nano 6, 6730–6740 (2012).
- Sharifi, S., Caracciolo, G. & Mahmoudi, M. Biomolecular corona affects controlled release of drug payloads from nanocarriers. *Trends Pharmacol. Sci.* 41, 641–652 (2020).
- Mahmoudi, M. The need for robust characterization of nanomaterials for nanomedicine applications. *Nat. Commun.* 12, 526 (2021).
 This comment emphasizes the necessity of standardization in protein corona research

to develop more efficient nanomedicines. Checklists, such as MIRIBEL (Minimum Information Reporting in Bio-Nano Experimental Literature), can improve the reliability and robustness of nanomedicine research.

- Zimmerman, L. J., Li, M., Yarbrough, W. G., Slebos, R. J. C. & Liebler, D. C. Global stability of plasma proteomes for mass spectrometry-based analyses. *Mol. Cell. Proteom.* 11, M111.014340 (2012).
- Ostroff, R. et al. The stability of the circulating human proteome to variations in sample collection and handling procedures measured with an aptamer-based proteomics array. *J. Proteom.* **73**, 649–666 (2010).
- 121. Gibb, B. C. Reproducibility. Nat. Chem. 6, 653-654 (2014).
- Liu, Y. et al. Multi-omic measurements of heterogeneity in HeLa cells across laboratories. Nat. Biotechnol. 37, 314–322 (2019).

- 123. Ashkarran, A. et al. Measurements of heterogeneity in proteomics analysis of nanoparticle protein corona across core facilities. Nat. Commun. 13, 6610 (2022). Variations in the protein corona analysis are investigated by sending identical corona samples to several proteomics core facilities and analysing resultant LC-MS/MS data. Although data correlated well between core facilities, the authors observed considerable heterogeneity. This study supports the need to develop protocols that improve protein corona characterization.
- Bondarenko, O. et al. Nanotoxicology and nanomedicine: the Yin and Yang of nano-bio interactions for the new decade. Nano Today 39, 101184 (2021).
- Westmeier, D., Stauber, R. H. & Docter, D. The concept of bio-corona in modulating the toxicity of engineered nanomaterials (ENM). *Toxicol. Appl. Pharmacol.* 299, 53–57 (2016).
- 126. Sharifi, S., Mahmoud, N. N., Voke, E., Landry, M. P. & Mahmoudi, M. Importance of standardizing analytical characterization methodology for improved reliability of the nanomedicine literature. *Nano Micro Lett.* **14**, 172 (2022). This perspective highlights how standardized protocols for protein corona analysis can improve NP-biological interface data reproducibility and clinical translation of nanotechnologies. Implementation of standardized protocols for protein corona research involves several stakeholders including research institutions, trainees and funding agencies.
- 127. Breiman, L. Random forests, Mach. Learn. 45, 5-32 (2001).
- Findlay, M. R., Freitas, D. N., Mobed-Miremadi, M. & Wheeler, K. E. Machine learning provides predictive analysis into silver nanoparticle protein corona formation from physicochemical properties. *Environ. Sci. Nano* 5, 64–71 (2018).

Machine learning is used to predict protein corona formation using few NP physicochemical properties and several protein characteristics as features. This study finds that protein features, and not NP physicochemical features, are more influential in predicting protein corona formation.

- Duan, Y. et al. Prediction of protein corona on nanomaterials by machine learning using novel descriptors. NanoImpact 17, 100207 (2020).
- Fluorescamine-labelled protein coronas are utilized as unique NP features to build random forest classifiers that could predict protein corona formation. These unique NP features are able to predict protein corona formation on 2D heterogeneous nanomaterials.
- Ban, Z. et al. Machine learning predicts the functional composition of the protein corona and the cellular recognition of nanoparticles. Proc. Natl Acad. Sci. USA 117, 10492–10499 (2020).

In this study, machine learning is used to predict functional proteins on NP protein coronas that are correlated with cellular associations. Protein corona composition is also predicted, and NP surface functionalization is found to be the most important factor that influenced the predictive power of the model.

- Walkey, C. D. et al. Protein corona fingerprinting predicts the cellular interaction of gold and silver nanoparticles. ACS Nano 8, 2439–2455 (2014).
- Liu, R., Jiang, W., Walkey, C. D., Chan, W. C. W. & Cohen, Y. Prediction of nanoparticlescell association based on corona proteins and physicochemical properties. *Nanoscale* 7, 9664–9675 (2015).
- Lazarovits, J. et al. Supervised learning and mass spectrometry predicts the in vivo fate of nanomaterials. ACS Nano 13, 8023–8034 (2019).
- Fourches, D. et al. Quantitative nanostructure-activity relationship modeling. ACS Nano 4, 5703–5712 (2010).
- Yamankurt, G. et al. Exploration of the nanomedicine-design space with high-throughput screening and machine learning. *Nat. Biomed. Eng.* 3, 318–327 (2019).
- Jumper, J. et al. Highly accurate protein structure prediction with AlphaFold. Nature 596, 583–589 (2021).
- Varadi, M. et al. AlphaFold Protein Structure Database: massively expanding the structural coverage of protein-sequence space with high-accuracy models. *Nucleic Acids Res.* 50, D439–D444 (2022).
- Li, G. et al. Coronavirus infections and immune responses. J. Med. Virol. 92, 424–432 (2020).
- Igakura, T. et al. Spread of HTLV-I between lymphocytes by virus-induced polarization of the cytoskeleton. Science 299, 1713–1716 (2003).
- Merad, M., Blish, C. A., Sallusto, F. & Iwasaki, A. The immunology and immunopathology of COVID-19. Science 375, 1122–1127 (2022).
- Akinc, A. et al. The Onpattro story and the clinical translation of nanomedicines containing nucleic acid-based drugs. *Nat. Nanotechnol.* 14, 1084–1087 (2019).
- Joshi, A. S., Zhang, H. & Prinz, W. A. Organelle biogenesis in the endoplasmic reticulum. Nat. Cell Biol. 19, 876–882 (2017).
- Shpilka, T. & Haynes, C. M. The mitochondrial UPR: mechanisms, physiological functions and implications in ageing. Nat. Rev. Mol. Cell Biol. 19, 109–120 (2018).
- 144. Behzadi, S. et al. Cellular uptake of nanoparticles: journey inside the cell. Chem. Soc. Rev. 46, 4218–4244 (2017).
- Hajipour, M. J. et al. Sex as an important factor in nanomedicine. Nat. Commun. 12, 2984 (2021).
- Bartleson, J. M. et al. SARS-CoV-2, COVID-19 and the aging immune system. Nat. Aging 1, 769–782 (2021).
- 147. Takahashi, T. et al. Sex differences in immune responses that underlie COVID-19 disease outcomes. *Nature* **588**, 315–320 (2020).
- Lundqvist, M. et al. The evolution of the protein corona around nanoparticles: a test study. ACS Nano 5, 7503–7509 (2011).

- 149. Schleh, C. et al. Size and surface charge of gold nanoparticles determine absorption across intestinal barriers and accumulation in secondary target organs after oral administration. Nanotoxicology 6, 36–46 (2012).
- Papafilippou, L., Claxton, A., Dark, P., Kostarelos, K. & Hadjidemetriou, M. Nanotools for sepsis diagnosis and treatment. Adv. Healthc. Mater. 10, e2001378 (2021).
- Digiacomo, L. et al. Efficient pancreatic cancer detection through personalized protein corona of gold nanoparticles. *Biointerphases* 16, 011010 (2021).
- Barui, A. K., Oh, J. Y., Jana, B., Kim, C. & Ryu, J. Cancer-targeted nanomedicine: overcoming the barrier of the protein corona. Adv. Ther. 3, 1900124 (2020).
- 153. Ju, Y. et al. Person-specific biomolecular coronas modulate nanoparticle interactions with immune cells in human blood. ACS Nano 14, 15723–15737 (2020).
- Aguado, B. A., Grim, J. C., Rosales, A. M., Watson-Capps, J. J. & Anseth, K. S. Engineering precision biomaterials for personalized medicine. Sci. Transl Med. 10, eaam8645 (2018).
- Lazarovits, J. et al. Synthesis of patient-specific nanomaterials. Nano Lett. 19, 116–123 (2019).
- Atkinson, S. P., Andreu, Z. & Vicent, M. J. Polymer therapeutics: biomarkers and new approaches for personalized cancer treatment. J. Pers. Med. 8, 6 (2018).
- Buck, L. B. & Axel, R. A novel multigene family may encode odorant receptors: a molecular basis for odor recognition. *Cell* 65, 175–187 (1991).
- Askim, J. R., Mahmoudi, M. & Suslick, K. S. Optical sensor arrays for chemical sensing: the optoelectronic nose. *Chem. Soc. Rev.* 42, 8649–8682 (2013).
- Mahmoudi, M., Lohse, S. E., Murphy, C. J. & Suslick, K. S. Identification of nanoparticles with a colorimetric sensor array. ACS Sens. 1, 17–21 (2016).
- Lin, Y. T. et al. Global plasma metabolomics to identify potential biomarkers of blood pressure progression. Arterioscler. Thromb. Vasc. Biol. 40, E227–E237 (2020).
- Csete, M. & Doyle, J. Bow ties, metabolism and disease. Trends Biotechnol. 22, 446–450 (2004).
- Tavakol, M. et al. Disease-related metabolites affect protein–nanoparticle interactions. Nanoscale 10, 7108–7115 (2018).
- Palchetti, S. et al. Effect of glucose on liposome-plasma protein interactions: relevance for the physiological response of clinically approved liposomal formulations. *Adv. Biosyst.* 3, e1800221 (2019).
- Maurer-Jones, M. A., Gunsolus, I. L., Murphy, C. J. & Haynes, C. L. Toxicity of engineered nanoparticles in the environment. *Anal. Chem.* 85, 3036–3049 (2013).
- Mueller, N. C. & Nowack, B. Exposure modeling of engineered nanoparticles in the environment. *Environ. Sci. Technol.* 42, 4447–4453 (2008).
- 166. Shahul Hamid, F. et al. Worldwide distribution and abundance of microplastic: how dire is the situation? Waste Manag. Res. 36, 873–897 (2018).
- Demirer, G. S. et al. High aspect ratio nanomaterials enable delivery of functional genetic material without DNA integration in mature plants. *Nat. Nanotechnol.* 14, 456–464 (2019).
- 168. Zhang, H. et al. Nanoparticle cellular internalization is not required for RNA delivery to mature plant leaves. *Nat. Nanotechnol.* **17**, 197–205 (2022).
- 169. Thabet, A. F. et al. Silica nanoparticles as pesticide against insects of different feeding types and their non-target attraction of predators. Sci. Rep. 11, 14484 (2021).
- Wang, D. et al. Nano-enabled pesticides for sustainable agriculture and global food security. Nat. Nanotechnol. 17, 347–360 (2022).
- 171. Nasser, F. & Lynch, I. Secreted protein eco-corona mediates uptake and impacts of polystyrene nanoparticles on *Daphnia magna*. J. Proteom. **137**, 45–51 (2016).
- Ong, K. J. et al. Humic acid ameliorates nanoparticle-induced developmental toxicity in zebrafish. *Environ. Sci. Nano* 4, 127–137 (2017).
- Fadare, O. O. et al. Humic acid alleviates the toxicity of polystyrene nanoplastic particles to: Daphnia magna. Environ. Sci. Nano 6, 1466–1477 (2019).
- Lipczynska-Kochany, E. Humic substances, their microbial interactions and effects on biological transformations of organic pollutants in water and soil: a review. *Chemosphere* 202, 420–437 (2018).
- Liu, F. et al. Modulation of cell uptake and cytotoxicity by nanoparticles with various physicochemical properties after humic acid adsorption. *Environ. Sci. Nano* 8, 3746–3761 (2021).
- Markiewicz, M. et al. Changing environments and biomolecule coronas: consequences and challenges for the design of environmentally acceptable engineered nanoparticles. *Green. Chem.* 20, 4133–4168 (2018).
- Radic, S. et al. Competitive binding of natural amphiphiles with graphene derivatives. Sci. Rep. 3, 2273 (2013).
- Xu, L. et al. The crucial role of environmental coronas in determining the biological effects of engineered nanomaterials. Small 16, e2003691 (2020).
- 179. Rummel, C. D., Jahnke, A., Gorokhova, E., Kühnel, D. & Schmitt-Jansen, M. Impacts of biofilm formation on the fate and potential effects of microplastic in the aquatic environment. *Environ. Sci. Technol. Lett.* 4, 258–267 (2017).
- Wang, Z., Zhang, L., Zhao, J. & Xing, B. Environmental processes and toxicity of metallic nanoparticles in aquatic systems as affected by natural organic matter. *Environ. Sci. Nano* 3, 240–255 (2016).
- Ekvall, M. T. et al. Adsorption of bio-organic eco-corona molecules reduces the toxic response to metallic nanoparticles in *Daphnia magna*. Sci. Rep. 11, 10784 (2021).
- 182. Gunsolus, I. L., Mousavi, M. P. S., Hussein, K., Bühlmann, P. & Haynes, C. L. Effects of humic and fulvic acids on silver nanoparticle stability, dissolution, and toxicity. *Environ. Sci. Technol.* 49, 8078–8086 (2015).
- Voke, E., Pinals, R. L., Goh, N. S. & Landry, M. P. In planta nanosensors: understanding biocorona formation for functional design. ACS Sens. 6, 2802–2814 (2021).

- Lehner, R., Weder, C., Petri-Fink, A. & Rothen-Rutishauser, B. Emergence of nanoplastic in the environment and possible impact on human health. *Environ. Sci. Technol.* 53, 1748–1765 (2019).
- Buchman, J. T., Hudson-Smith, N. V., Landy, K. M. & Haynes, C. L. Understanding nanoparticle toxicity mechanisms to inform redesign strategies to reduce environmental impact. Acc. Chem. Res. 52, 1632–1642 (2019).
- Priester, J. H. et al. Soybean susceptibility to manufactured nanomaterials with evidence for food quality and soil fertility interruption. *Proc. Natl Acad. Sci. USA* **109**, 2451–2456 (2012).
- Zhu, H., Han, J., Xiao, J. Q. & Jin, Y. Uptake, translocation, and accumulation of manufactured iron oxide nanoparticles by pumpkin plants. J. Environ. Monit. 10, 713–717 (2008).
- Lin, D. & Xing, B. Phytotoxicity of nanoparticles: inhibition of seed germination and root growth. Environ. Pollut. 150, 243–250 (2007).
- Thiele, C. J., Hudson, M. D., Russell, A. E., Saluveer, M. & Sidaoui-Haddad, G. Microplastics in fish and fishmeal: an emerging environmental challenge? Sci. Rep. 11, 2045 (2021).
- 190. Leslie, H. A. et al. Discovery and quantification of plastic particle pollution in human blood. *Environ. Int.* **163**, 107199 (2022).
- Zhang, N., Li, Y. B., He, H. R., Zhang, J. F. & Ma, G. S. You are what you eat: microplastics in the feces of young men living in Beijing. Sci. Total Environ. 767, 144345 (2021).
- Farvardin, A. et al. The apoplast: a key player in plant survival. Antioxidants 9, 604 (2020).
 Faria, M., Björnmalm, M., Crampin, E. J. & Caruso, F. A few clarifications on MIRIBEL. Nat. Nanotechnol. 15, 2–3 (2020).
- Faria, M. et al. Minimum information reporting in bio-nano experimental literature. Nat. Nanotechnol. 13, 777–785 (2018).
- Lammers, T. & Storm, G. Setting standards to promote progress in bio-nano science. Nat. Nanotechnol. 14, 626 (2019).
- Schoenmaker, L. et al. mRNA-lipid nanoparticle COVID-19 vaccines: structure and stability. Int. J. Pharm. 601, 120586 (2021).
- Nance, E., Pun, S. H., Saigal, R. & Sellers, D. L. Drug delivery to the central nervous system. Nat. Rev. Mater. 7, 314–331 (2022).
- Liu, R. & Lal, R. Potentials of engineered nanoparticles as fertilizers for increasing agronomic productions. Sci. Total Environ. 514, 131–139 (2015).
- Wang, L. et al. Silica nanoparticles activate defense responses by reducing reactive oxygen species under *Ralstonia solanacearum* infection in tomato plants. *NanoImpact* 28, 100418 (2022).
- 200. Keshishian, H. et al. Quantitative, multiplexed workflow for deep analysis of human blood plasma and biomarker discovery by mass spectrometry. *Nat. Protoc.* 12, 1683–1701 (2017).
- Etzioni, R. et al. The case for early detection. Nat. Rev. Cancer 3, 243–252 (2003).
 Cuzick, J. et al. Prevention and early detection of prostate cancer. Lancet Oncol. 15, 15
- e484-e492 (2014).
- Moman, R. N., Gupta, N. & Varacalla, M. Physiology, Albumin (StatPearls, 2022).
 Geyer, P. E., Holdt, L. M., Teupser, D. & Mann, M. Revisiting biomarker discovery by plasma
- proteomics. *Mol.* Syst. *Biol.* **13**, 942 (2017). 205. Jacobs, J. M. et al. Utilizing human blood plasma for proteomic biomarker discovery.
- J. Proteome Res. 4, 1073–1085 (2005). 206. Stastna, M. & Van Eyk, J. E. Secreted proteins as a fundamental source for biomarker discussion of Detection 27 270 (2012).
- discovery. Proteomics 12, 722–735 (2012).
 207. Mann, M., Kulak, N. A., Nagaraj, N. & Cox, J. The coming age of complete, accurate, and ubiquitous proteomes. *Mol. Cell* 49, 583–590 (2013).
- Saavedra, J., Stoll, S. & Slaveykova, V. I. Influence of nanoplastic surface charge on eco-corona formation, aggregation and toxicity to freshwater zooplankton. *Environ. Pollut.* 252, 715–722 (2019).
- Baalousha, M., Manciulea, A., Cumberland, S., Kendall, K. & Lead, J. R. Aggregation and surface properties of iron oxide nanoparticles: influence of pH and natural organic matter. *Environ. Toxicol. Chem.* 27, 1875–1882 (2008).

- Partikel, K., Korte, R., Mulac, D., Humpf, H. U. & Langer, K. Serum type and concentration both affect the protein-corona composition of PLGA nanoparticles. *Beilstein J. Nanotechnol.* 10, 1002–1015 (2019).
- Cai, R. et al. Dynamic intracellular exchange of nanomaterials' protein corona perturbs proteostasis and remodels cell metabolism. Proc. Natl Acad. Sci. USA 119, e2200363119 (2022).
- Albanese, A. et al. Secreted biomolecules alter the biological identity and cellular interactions of nanoparticles. ACS Nano 8, 5515–5526 (2014).
- Briffa, S. M., Nasser, F., Valsami-Jones, E. & Lynch, I. Uptake and impacts of polyvinylpyrrolidone (PVP) capped metal oxide nanoparticles on *Daphnia magna*: role of core composition and acquired corona. *Environ. Sci. Nano* 5, 1745–1756 (2018).
- Grintzalis, K., Lawson, T. N., Nasser, F., Lynch, I. & Viant, M. R. Metabolomic method to detect a metabolite corona on amino-functionalized polystyrene nanoparticles. *Nanotoxicology* 13, 783–794 (2019).
- Galloway, T. S., Cole, M. & Lewis, C. Interactions of microplastic debris throughout the marine ecosystem. Nat. Ecol. Evol. 1, 0116 (2017).

Acknowledgements

The authors acknowledge support of the US National Institute of Diabetes and Digestive and Kidney Diseases (grant DK131417) (to M.M.), a Burroughs Wellcome Fund Career Award at the Scientific Interface (to M.P.L.), a Dreyfus Foundation Award (to M.P.L.), the Philomathia Foundation (to M.P.L.), an NIH MRA Award R35GM128922 (to M.P.L.), an NIH R21 NIDA Award 1R03DA052810 (to M.P.L.), an NIH RA Award R35GM128922 (to M.P.L.), an NSF CBET Award 1733575 (to M.P.L.), a CZI Imaging Award (to M.P.L.), a Sloan Foundation Award (to M.P.L.), a USDA BBT EAGER Award (to M.P.L.), a More Foundation Award (to M.P.L.), a USDA BBT EAGER Award (to M.P.L.), a More Foundation Award (to M.P.L.), a DC Office of Science grant DE-SC0020366 (to M.P.L.). M.P.L. is a Chan Zuckerberg Biohub investigator, a Hellen Wills Neuroscience Institute investigator and an IGI investigator.

Author contributions

All the authors researched data for the article, contributed substantially to discussion of the content, wrote the article and reviewed and/or edited the manuscript before submission.

Competing interests

M.M. discloses that he is an author for patent US17/215,923 — system and method for protein corona sensor array for early detection of diseases. The other authors declare no competing interests.

Additional information

Peer review information Nature Reviews Materials thanks Pu Chun Ke and the other, anonymous, reviewer(s) for their contribution to the peer review of this work.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author selfarchiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

Related links

AlphaFold: https://alphafold.ebi.ac.uk/ Golestan cohort: https://dceg2.cancer.gov/gemshare/studies/GCS/

© Springer Nature Limited 2023